

Accession # 00280395

Female Sample Report 123 A Street Sometown, CA 90266



Last Menstrual Period:

Ordering Physician:

Precision Analytical

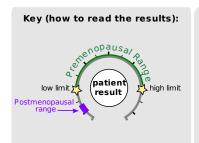
DOB: 1976-01-01

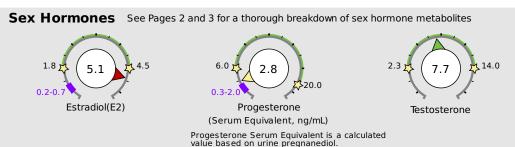
Age: 41

Gender: Female

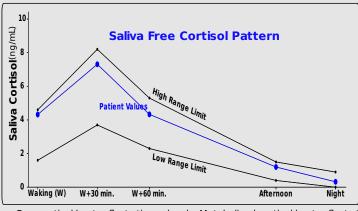
Collection Times:
2017-04-14 06:05AM (U)
2017-04-14 08:00AM (U)
2017-04-14 05:05PM (U)
2017-04-14 10:10PM (U)
2017-04-14 06:00AM (S)
2017-04-14 06:30AM (S)
2017-04-14 07:00AM (S)
2017-04-14 05:00PM (S)
2017-04-14 10:00PM (S)

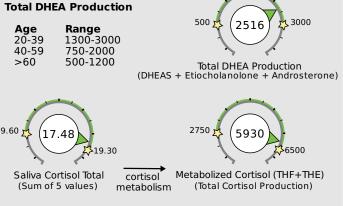
Hormone Testing Summary





Adrenal Hormones See pages 4 and 5 for a more complete breakdown of adrenal hormones





Free cortisol best reflects tissue levels. Metabolized cortisol best reflects total cortisol production.

The following videos (which can also be found on the website under the listed names along with others) may aid your understanding:

<u>DUTCH Plus Overview</u> (quick overview) <u>Estrogen Tutorial</u> <u>Female Androgen Tutorial</u> <u>Cortisol/CAR Tutorial</u>

PLEASE BE SURE TO READ BELOW FOR ANY SPECIFIC LAB COMMENTS. More detailed comments can be found on page 8.

- The patient collected an "Insomnia" salivary sample in the middle of the night. The cortisol result for this sample was 2.10ng/mL (expected range 0-0.9). Please see page 4 for cortisol and cortisone results for this sample.

The Cortisol Awakening Response (CAR) was 2.99ng/mL (expected range 1.5-4.0) or 69.2% (range 50-160%). See page 5 for more details.



Accession # 00280395 Female Sample Report 123 A Street Sometown, CA 90266



Sex Hormones and Metabolites

Ordering Physician: Precision Analytical

DOB: 1976-01-01

Age: 41

Gender: Female

Last Menstrual Period:

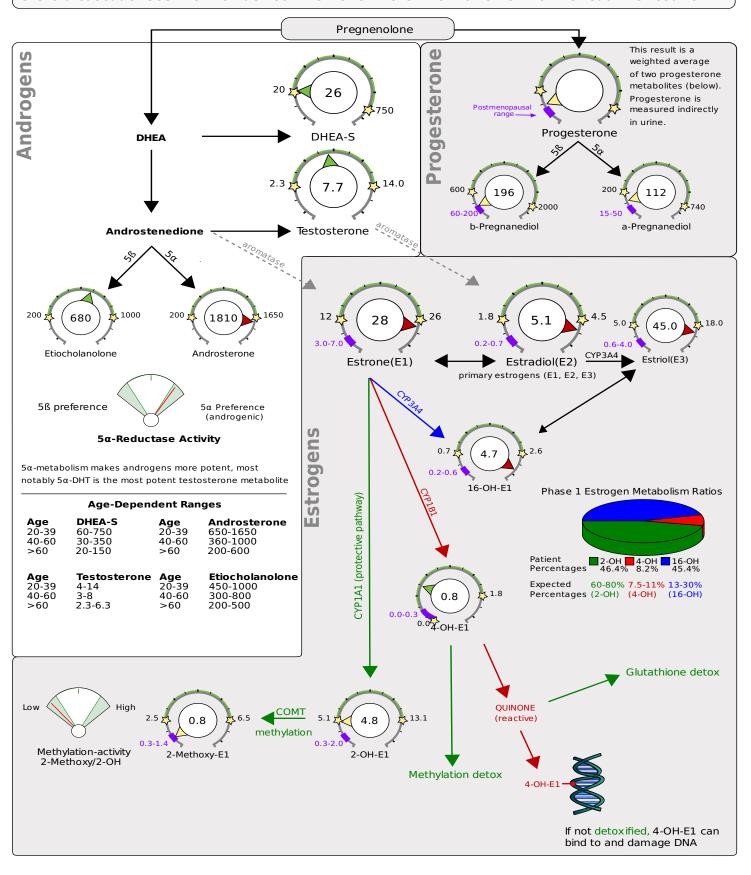
Collection Times:
2017-04-14 06:05AM (U)
2017-04-14 08:00AM (U)
2017-04-14 05:05PM (U)
2017-04-14 10:10PM (U)
2017-04-14 06:30AM (S)
2017-04-14 07:00AM (S)
2017-04-14 07:00AM (S)
2017-04-14 07:00AM (S)
2017-04-14 07:00AM (S)

Test		Result	Units	Luteal*	Postmenopausal
Progesterone M	etabolites (Urine)			Range	Range
b-Pregnanediol	Below luteal range	196.0	ng/mg	600 - 2000	60-200
a-Pregnanediol	Below luteal range	112.0	ng/mg	200 - 740	15-50
Estrogens and M	1etabolites (Urine)				
Estrone(E1)	Above luteal range	28.2	ng/mg	12 - 26	3.0-7.0
Estradiol(E2)	Above luteal range	5.1	ng/mg	1.8 - 4.5	0.2-0.7
Estriol(E3)	Above luteal range	45.0	ng/mg	5 - 18	0.6-4.0
2-OH-E1	Below luteal range	4.8	ng/mg	5.1 - 13.1	0.3-2.0
4-OH-E1	Within luteal range	8.0	ng/mg	0 - 1.8	0-0.3
16-OH-E1	Above luteal range	4.7	ng/mg	0.7 - 2.6	0.2-0.6
2-Methoxy-E1	Below luteal range	8.0	ng/mg	2.5 - 6.5	0.3-1.4
2-OH-E2	Low end of luteal range	0.19	ng/mg	0 - 1.2	0-0.3
4-OH-E2	Within luteal range	0.2	ng/mg	0 - 0.5	0-0.1
2-Methoxy-E2	Within luteal range	0.4	ng/mg	0 - 0.7	0-0.4
Total Estrogen	Above range	89.79	ng/mg	35 - 70	4.0-15
Androgens and	Metabolites (Urine)				
DHEA-S	Low end of range	26.0	ng/mg	20 - 750	
Androsterone	Above range	1810.0	ng/mg	200 - 1650	
Etiocholanolone	Within range	680.0	ng/mg	200 - 1000	
Testosterone	Within range	7.7	ng/mg	2.3 - 14	
5a-DHT	Above range	7.2	ng/mg	0 - 6.6	
5a-Androstanediol	Above range	42.0	ng/mg	12 - 30	
5b-Androstanediol	Within range	32.0	ng/mg	20 - 75	
Epi-Testosterone	Within range	8.8	ng/mg	2.3 - 14	

*the Luteal Range is the premenopausal range. When patients are taking oral progesterone this range for progesterone metabolites is not luteal and reflects the higher levels expected when patients take oral progesterone. This test is intended to be taken in the luteal phase of the menstrual cycle (days 19-22 of a 28 day cycle) for premenopausal women. The ranges in the table below may be used when samples are taken during the first few days (follicular) of the cycle, during ovulation (days 11-14) or when patients are on oral progesterone. See the following pages for age-dependent ranges for androgen metabolites.

Additional Normal Ranges	Follicular	Ovulatory	Oral Pg (100mg)
b-Pregnanediol	100-300	100-300	2000-9000
a-Pregnanediol	25-100	25-100	580-3000
Estrone (E1)	4.0-12.0	22-68	N/A
Estradiol (E2)	1.0-2.0	4.0-12.0	N/A

Hormone metabolite results from the previous page are presented here as they are found in the steroid cascade. See the Provider Comments for more information on how to read the results.





Accession # 00280395 Female Sample Report 123 A Street Sometown, CA 90266



Adrenal

Ordering Physician: Precision Analytical

DOB: 1976-01-01

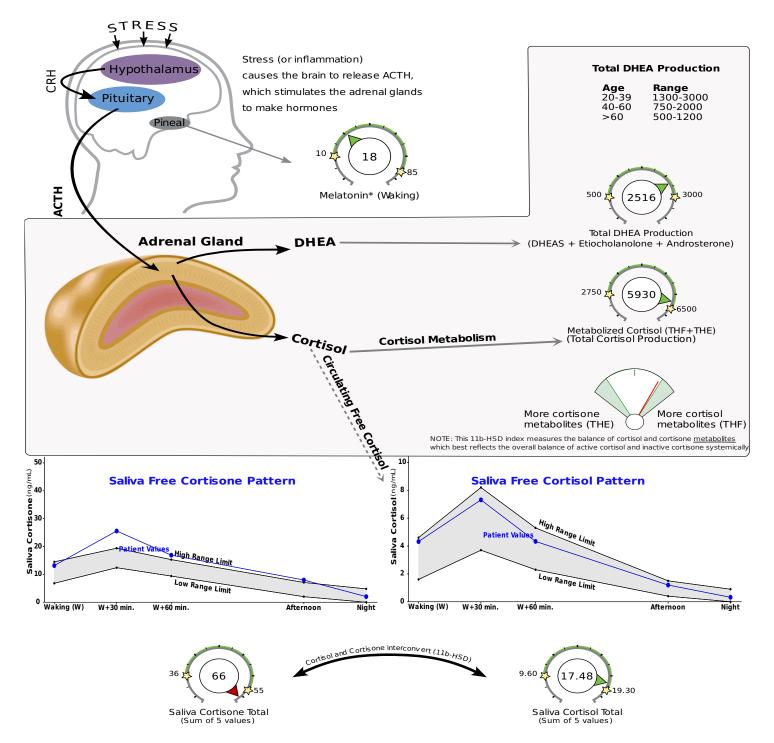
Age: 41

Gender: Female

Last Menstrual Period:

Collection Times:
2017-04-14 06:05AM (U)
2017-04-14 08:00AM (U)
2017-04-14 05:05PM (U)
2017-04-14 10:10PM (U)
2017-04-14 06:00AM (S)
2017-04-14 06:30AM (S)
2017-04-14 07:00AM (S)
2017-04-14 05:00PM (S)
2017-04-14 10:00PM (S)
2017-04-14 01:30AM (S*)

Category	Test		Result	Units	Normal Range
	sol and Cortisone (Saliva)				
	Saliva Cortisol - Waking (W)	High end of range	4.32	ng/mL	1.6 - 4.6
	Saliva Cortisol - W+30 min.	High end of range	7.31	ng/mL	3.7 - 8.2
	Saliva Cortisol - W+60 min.	Within range	4.33	ng/mL	2.3 - 5.3
	Saliva Cortisol - Afternoon	Within range	1.2	ng/mL	0.4 - 1.5
	Saliva Cortisol - Night	Within range	0.32	ng/mL	0 - 0.9
	Saliva Cortisone - Waking (W)	High end of range	13.16	ng/mL	6.8 - 14.5
	Saliva Cortisone - W+30 min.	Above range	25.57	ng/mL	12.4 - 19.4
	Saliva Cortisone - W+60 min.	Above range	16.9	ng/mL	9.4 - 15.3
	Saliva Cortisone - Afternoon	Above range	7.98	ng/mL	2 - 7.1
	Saliva Cortisone - Night	Within range	2.02	ng/mL	0 - 4.8
	Saliva Cortisol Total	High end of range	17.48	ng/mL	9.6 - 19.3
	Saliva Cortisone Total	Above range	65.63	ng/mL	36 - 55
Creatinine	(Urine)				
	Creatinine A (Waking)	Within range	0.4	mg/ml	0.2 - 2
	Creatinine B (Morning)	Within range	0.51	mg/ml	0.2 - 2
	Creatinine C (Afternoon)	Within range	0.92	mg/ml	0.2 - 2
	Creatinine D (Night)	Within range	1.01	mg/ml	0.2 - 2
Cortisol Mo	etabolites and DHEA-S (Urine)				
	a-Tetrahydrocortisol (a-THF)	Above range	400.0	ng/mg	75 - 370
	b-Tetrahydrocortisol (b-THF)	Above range	2500.0	ng/mg	1050 - 2500
	b-Tetrahydrocortisone (b-THE)	Within range	3030.0	ng/mg	1550 - 3800
	Metabolized Cortisol (THF+THE)	High end of range	5930.0	ng/mg	2750 - 6500
	DHEA-S	Low end of range	26.0	ng/mg	20 - 750
Additional	Cortisol and Cortisone (Saliva)				
	Saliva Cortisol - Insomnia	Above range	2.1	ng/mL	0 - 0.9
k	Saliva Cortisone - Insomnia	Above range	10.4	ng/mL	0 - 4.8



- The patient submitted an Insomnia salivary sample. The cortisol result for this sample was 2.10ng/mL (expected range 0-0.9) The cortisone result for this sample was 10.4 ng/mL (expected range 0-4.8)

The Cortisol Awakening Response (CAR) is the rise in salivary cortisol between the waking sample and the sample collected 30 (as well as 60) minutes later. This "awakening response" is essentially a "mini stress test" and is a useful measurement in addition to the overall up-and-down (diurnal) pattern of free cortisol throughout the day. This patient shows a waking cortisol of 4.32 and an increase to 7.3 after 30.0 minutes. This is an increase of 2.99ng/mL or 69.2%. Expected increases differ depending on the methods used. Preliminary research shows that 50-160% or 1.5-4.0ng/mL increases are common with samples collected 30 minutes after waking. These guidelines are considered research only.

This patient shows a salivary cortisol of 4.33 measured 60 minutes after waking. This is an increase of 0.01ng/mL or 0.23% compared to the waking sampe. To date, data suggests that expected results may be 0-70%, and this guideline is considered for research only.



Accession # 00280395 Female Sample Report 123 A Street Sometown, CA 90266



Organic Acid Tests (OATs)

Ordering Physician: Precision Analytical

DOB: 1976-01-01

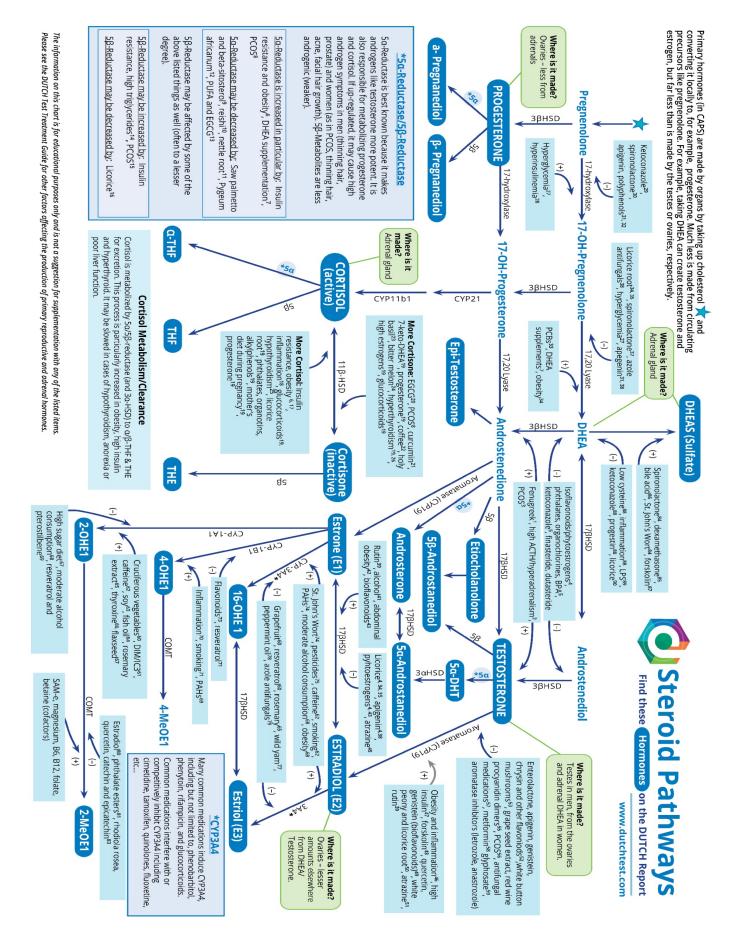
Age: 41

Gender: Female

Last Menstrual Period:

Collection Times:
2017-04-14 06:05AM (U)
2017-04-14 08:00AM (U)
2017-04-14 05:05PM (U)
2017-04-14 10:10PM (U)
2017-04-14 06:00AM (S)
2017-04-14 07:00AM (S)
2017-04-14 07:00AM (S)
2017-04-14 07:00AM (S)
2017-04-14 10:00PM (S)

Category	Test		Result	Units	Normal Range				
	Nutritional Organic Acids								
Vitamin B12	Marker (may be deficient if high	n) - (Urine)							
	Methylmalonate (MMA)	Within range	1.2	ug/mg	0 - 2.2				
Vitamin B6 M	larkers (may be deficient if high	ı) - (Urine)							
	Xanthurenate	Above range	6.8	ug/mg	0 - 1.4				
	Kynurenate	Above range	35.5	ug/mg	0 - 7.3				
Glutathione N	Marker (may be deficient if low o	or high) - (Urine)							
	Pyroglutamate	Below range	23.2	ug/mg	32 - 60				
	Neur	otransmitter Metabo	lites						
Dopamine M	etabolite - (Urine)								
	Homovanillate (HVA)	Low end of range	5.6	ug/mg	4 - 13				
Norepinephri	ne/Epinephrine Metabolite - (Ur	ine)							
	Vanilmandelate (VMA)	Within range	4.8	ug/mg	2.4 - 6.4				
Melatonin (*r	Melatonin (*measured as 6-OH-Melatonin-Sulfate) - (Urine)								
	Melatonin* (Waking)	Low end of range	18.2	ng/mg	10 - 85				
Oxidative Str	Oxidative Stress / DNA Damage, measured as 8-Hydroxy-2-deoxyguanosine (8-OHdG) - (Urine)								
	8-OHdG (Waking)	High end of range	4.3	ng/mg	0 - 5.2				



References

- Biochem, 2010. **116**(3): p. 146-55. Simonian, M.H., ACTH and thyroid hormone regulation of 3 be-Hamden, K., et al., Potential protective effect on key steroidogen sis and metabolic enzymes and sperm abnormalities by fenugreek steroids in testis and epididymis of surviving diabetic rats. *Arch Physiol*
- Kaaijk, E.M., et al., Distribution of steroidogenic enzymes involved cortical cells. J Steroid Biochem, 1986. 25(6): p. 1001-6. ta-hydroxysteroid dehydrogenase activity in human fetal adreno-
- lizing enzymes. J Steroid Biochem Mol Biol, 2005. 93(2-5); p. 285-92. Zhang, S., et al., Endocrine disruptors of inhibiting testicular 38-hydroxysteroid dehydrogenase. Chem Biol Interact, 2019. 303: in androgen synthesis in polycystic ovaries: an immunohistochem cal study. Mol Hum Reprod, 2000. 6(5): p. 443-7. D., et al., Inhibition of 17beta-hydroxysteroid dehydroge-
- dehydrogenase type 1 expression and elevated hepatic 5alpha-re Tomlinson, J.W., et al., Impaired glucose tolerance and insulin resis tance are associated with increased adipose 11beta-hydroxysteroic activity. Diabetes, 2008. 57(10): p. 2652-60.
- Stomati, M., et al., Six-month oral dehydroepiandrosterone supplementation in early and late postmenopause. *Gynecol Endocri* 2000. **14**(5): p. 342-63. ation in early and late postmenopause. Gynecol Endocrino 30. 29.
- Prager, $N_{\cdot\cdot}$ et al., A randomized, double-blind, placebo-controlled trial to determine the effectiveness of botanically derived inhibitors rates. Tsilchorozidou, T., J.W. Honour, and G.S. Conway, Altered cortisol metabolism in polycystic ovary syndrome: insulin enhances Salpha-reduction but not the elevated adrenal steroid production J Clin Endocrinol Metab, 2003. **88**(12): p. 5907-13. 32. <u>ω</u>
- Fujita, R., et al., Anti-androgenic activities of Ganoderma lucidum. of 5-alpha-reductase in the treatment of androgenetic alopecia. J Altern Complement Med, 2002. 8(2): p. 143-52. 33
- Moradi, H.R., et al., The histological and histometrical effects of Urtica dioica extract on rat's prostate hyperplasia. Vet Res Forum, Ethnopharmacol, 2005. 102(1): p. 107-12.

= 10.

12.

35 34

13 2015. 6(1): p. 23-9.
Will, T., et al., Pygeum africanum for benign prostatic hyperplasia.
Cochrone Database Syst Rev, 2002(1): p. CD001044.
Azzouni, F., et al., The S alpha-reductase isozyme family: a review of basic biology and their role in human diseases. Adv Urol, 2012.

14.

- 2012: p. 530121.

 Westerbacka, J., et al., Body fat distribution and cortisol metabolism Westerbacka, J., et al., Body fat distribution and cortisol in healthy men: enhanced Speta-reductase and lower cortisol/ in healthy men: enhanced speta-reductase and lower cortisol/ in healthy men: enhanced speta-reductase and lower cortisol/ metabolite ratios in men with fatty liver. J Clin Endocrinol
- Metab. 2003. 88(10): p. 4924-31.
 Gambineri, A., et al., Increased clearance of cortisol by 5beta-reduction as in a subgroup of women with adrenal hyperandrogenism in polycystic ovary syndrome. J Endocrinol Invest. 2009. 32(3): p. 210-8.

 Ojima, M., et al., Ilm einhibitory effects of bytycrhizin and bytyr-rheinic acid on the metabolism of cortisol and prednisolone-in

39

vivo and in vitro studies]. Nihon Naibunpi Gakkai Zasshi, 1990. 66(5)

16. 15.

- activity in subcutaneous adipose tissue in humans: implications in obesity and diabetes. J Clin Endocrinol Metab, 2015. 100(1): p. E70-6 Dube, S., et al., 11β-hydroxysteroid dehydrogenase types 1 and 2
- droxysteroid dehydrogenase type 1 (11β-HSD1) in human adipo-cytes is mediated by MEK, C/EBPβ, and NF-κB/ReIA. J Clin Endocrinol Esteves, C.L., et al., Proinflammatory cytokine induction of 11β-hy Metab, 2014. 99(1): p. E160-8.

<u>.</u>8 17.

Chapman, K., M. Holmes, and J. Seckl, 11β-hydroxysteroid dehydro genases: intracellular gate-keepers of tissue glucocorticoid action. *Physiol Rev*, 2013. **93**(3): p. 1139-206.

19.

- 21 20. teroid dehydrogenase type 1. PLoS One, 2014. 9(1): p. e84468. catechine-3-gallate, are potent inhibitors of human 11β-h Hintzpeter, J., et al., Green tea and one of its constituents, Epigallo ydroxys-
- 11β-hydroxysteroid dehydrogenase 1: improving lipid profiles in high-fat-diet-treated rats. PLoS One, 2013. 8(3): p. e49976. Atanasov, A.G., et al., Coffee inhibits the reactivation of glucocorti Hu, G.X., et al., Curcumin as a potent and selective inhibitor of , et al., Coffee inhibits the reactivation of glucocorti

22

- Jothle Richard, E., et al., Anti-stress Activity of Ocimum sanctum: Possible Effects on Hypothalamic-Pituitary-Adrenal Axis. *Phytother*
- Res, 2016. 30(5): p. 805-14.

 Blum, A., et al., Momordica chrarantia extract, a herbal remedy for type 2 diabetes, contains a specific 11B-hydroxysteroid dehydrogenase type 1 inhibitor. J Steroid Biochem Mol Biol, 2012. 128(1-2): p.

24. 23.

- tabolites in hyperthyroid and hypothyroid patients. Clin Endocrinol Hoshiro, M., et al., Comprehensive study of urinary cortisol me-
- (0xf), 2006. 64(1); p. 37-45.
 Taniyama, M., K. Honma, and Y. Ban, Urinary cortisol metabolites in the assessment of peripheral thyroid hormone action: application for diagnosis of resistance to thyroid hormone. *Thyroid*, 1993. 3(3):

26. 25.

and increased 17-hydroxylase activities in type 2 diabetes mellitus Ueshiba, H., et al., Decreased steroidogenic enzyme 17,20-lyase

> 53. 52. 51. 50.

P450c17 alpha activity and serum free testosterone after reduction of insulin secretion in polycystic ovary syndrome. N Engl J Med. 1996. **335**(9): p. 617-23

42

- Endocrinol (Oxf), 1991. 35(2): p. 163-8. Kossor, D.C. and H.D. Colby, Dose-dependent actions of spironolac Clin
- hormones from human adrenocortical NCI-H295R cells. *Biol Pharm Bull*, 2013. **36**(2): p. 228-37.

57. 56. 55.

- Armanini, D., G. Bonanni, and M. Palermo, Reduction of serum testosterone in men by licorice. N Engl J Med, 1999. 341(15): p. 1158.
 Armanini, D., et al., Licorice reduces serum testosterone in healthy
- women. Steroids, 2004. **69**(11-12): p. 763-6. Serafini, P. and R.A. Lobo, The effects of spironolactone on adrenal

37. 36.

38.

- steroidogenesis in hirsute women. Fertil Steril. 1985. 44(5): p. 595-9. Ayub. M. and M.J. Levell. Inhibition of human adrenal steroidogenic enzymes in witro by imidazole drugs including ketoconazole. J Steroid Biochem. 1989. 32(4): p. 515-24. Wang. X., et al., Suppression of rat and human androgen biosynthetic enzymes by apligenin: Possible use for the treatment of prostate cancer. Filoteropia, 2016. 111: p. 66-72.

4.

40

42.

- Hu, T., et al., Brown adjoose tissue activation by rutin ameliorates polytystic ovary syndrome in rat. J Nutr Biochem, 2017, 47: p. 21-28. Sarkola, T., et al., Actue effect of alcohol on androgens in premenopausal women. Alcohol Alcohol, 2000. 35(1): p. 84-90. Corbould, A.M., et al., The effect of obesity on the ratio of typs 37 (7) beat-hydroxysteroid dehydrogenase mRNA to cytochrome P450 aromatase mRNA in subcutaneous abdominal and nura-abdominal aromatase mRNA in subcutaneous abdominal and nura-abdominal. adipose tissue of women. Int J Obes Relat Metab Disord, 2002. 26(2)
- p. 165-75. Krazeisen, A., et al., Human 17beta-hydroxysteroid dehydrogenase type 5 is inhibited by dietary flavonoids. Adv Exp Med Biol, 2002. 505: p. 151-61.
- Le Bail, J.C., et al., Effects of phytoestrogens on aromatase, 3beta and 17beta-hydroxysteroid dehydrogenase activities and human

4 43.

- 45 Abarikwu, S.O. and E.O. Farombi, Quercetin ameliorates atra-zine-induced changes in the testicular function of rats. *Toxicol* breast cancer cells. Life Sci, 2000. 66(14): p. 1281-91. Toxicol Ind
- 47. Health, 2016. **32**(7); p. 1278-85. Gérard, C. and K.A. Brown, Obesity and breast cancer - Role of es-
- Randolph, J.F., et al., The effect of insulin on aromatase activity in trogens and the molecular underpinnings of aromatase regulation in breast adipose tissue. *Mol Cell Endocrinol*, 2018. **466**: p. 15-30.

- 48.
- 49.

- Eur J Endocrinol, 2002. 146(3): p. 375-80.

 Nestler, J.E. and D.J. Jakubowicz, Decreases in ovarian cytochrome

28. 27.

- Engelhardt, D., et al., The influence of ketoconazole on human adrenal steroidogenesis: incubation studies with tissue slices.
- Pharmacology, 1992. **45**(1): p. 27-33.

 Hasegawa, E., et al., Effect of polyphenols on production of steroid tone on the inner and outer zones of the guinea pig adrenal cortex
- Marti, N., et al., Resveratrol inhibits androgen production of human adrenocortical H295R cells by lowering CYP17 and CYP21 expres-
- sion and activities. PLoS One. 2017. 12(3): p. e0174224. Andric, S.A., et al., Acute effects of polychlorinated biphenyl-containing and -free transformer fluids on rat testicular steroidogene-
- sis. Environ Health Perspect, 2000. 108(10): p. 955-9.
 Kim, S.H., et al., Body Fat Mass is Associated With Ratio of Steroid Metabolites Reflecting 17.20-Lyase Activity in Perspubertal Girls. J Clin Endocrinol Metab., 2016. 101(12): p. 4653-4660.

61.

- 63 62.
- 2 women. J Nutr., 2006. 136(6): p. 1588-95.

 Lu, L.J., et al., increased urinary secretion of 2-hydroxyestrone but not 16alpha-hydroxyestrone in premenopausal women during a soya diet containing isoflavones. Cancer Res, 2000. 60(5): p. 1299-305.

87. 86.

- Chen, H.W., et al., The combined effects of garlic oil and fish oil on the hepatic antioxidant and drug-metabolizing enzymes of rats, Br / Nutr, 2003, 89(2); p. 189-200.

 Debersac, P., et al., Induction of cytochrome P450 and/or detox-

88

- ine on C-2 and C-16 alpha hydroxylations of estradiol in humans. Steroids, 1990. S5(1); p. 22-6. Peters, L.P. and R.W. Teel. Effect of high sucrose diet on cyto-chrome P450 1 A and heterocyclic amine mutagenesis. *Anticance* Res, 2003. 23(1A); p. 399-403.
- tion on urinary estrogen metabolites in postmenopausal women i a controlled feeding study. Cancer Med, 2017. 6(10): p. 2419-2423. Licznerska, B., et al., Resveratrol and its methoxy derivatives mod-Mahabir, S., et al., Effects of low-to-moderate alcohol supplementa

69. 68 67. 66. 65.

70. Smerdová, L., et al., Upregulation of CYP1B1 expression by inflam matory cytokines is mediated by the p38 MAP kinase signal transmatory cytokines is mediated by the p38 MAP kinase sign duction pathway. Carcinogenesis, 2014. 35(11): p. 2534-43

- Watanabe, M. and S. Nakajin, Forskolin up-regulates aromatase (CYP19) activity and gene transcripts in the human adrenocortical carcinoma cell line H295R. J Endocrinol. 2004. 180(1): p. 15-33. Sanderson, J.T., et al., induction and hibition of aromatase (CYP19) activity by natural and synthetic flavonoid compounds in
- Takeuchi, T., et al., Effect of paeoniflorin, glycyrrhizin and glycyr-rhetic acid on ovarian androgen production. *Am J Chin Med*, 1991 H295R human adrenocortical carcinoma cells. Toxicol Sci, 2004
- ity in estrogen sensitive target tissues. J Appl Toxicol, 2008. 28(3): p. 260-70. Holloway, A.C., et al., Atrazine-induced changes in aromatase activ 19(1): p. 73-8.
- Lephart, E.D., Modulation of Aromatase by Phytoestrogens. Enzym Res, 2015. 2015: p. 594656
- Novaes, M.R., et al., The effects of dietary supplementation with Agaricales mushrooms and other medicinal fungi on breast cancer: evidence-based medicine. Clinics (Soo Paulo), 2011. 66(12): p. 2133-
- Eng. E.T., et al., Suppression of estrogen biosynthesis by procyani din dimers in red wine and grape seeds. Cancer Res, 2003. **63**(23) Satoh, K., et al., Inhibition of aromatase activity by green tea extrac in rats. Food Chem Toxicol, 2002. 40(7): p. 925-33. catechins and their endocrinological effects of oral administration
- Chen, J., et al., The correlation of aromatase activity and obesity in p. 8516-22 women with or without polycystic ovary syndrome. J Ovarian Res
- Ayub, M. and M.J. Levell, The inhibition of human prostatic aromatase activity by imidazole drugs including ketoconazole and 4-hydroxyandrostenedione. Biochem Pharmacol, 1990. 40(7): p. 2015. 8: p. 11.
- Rice, S., et al., Dual effect of metformin on growth inhibition and oestradiol production in breast cancer cells. *Int J Mol Med*, 2015.
- 35(4): p. 1088-94. Richard. S., et al., Differential effects of glyphosate and roundup on human placental cells and aromatase. *Environ Health Perspect*, 2005. 113(6): p. 716-20.
- Hodges, R.E. and D.M. Minich, Modulation of Metabolic Detoxifi-Scientific Review with Clinical Application. J Nutr Metab, 2015. 2015 cation Pathways Using Foods and Food-Derived Components: A

60. 59. 58.

Michnovicz, J.J., H. Adlercreutz, and H.L. Bradlow, Changes in levels of urinary estrogen metabolites after oral incide 3-carbinol treat-

84

- ment in humans. J Natl Concer Inst, 1997. **89**(10): p. 718-23. Sowers, M.R., et al., Selected diet and lifestyle factors are associated.
- ication enzymes by various extracts of rosemary; description of specific patterns. Food Chem Toxicol, 2001. **39**(9): p. 907-18. Michnovicz, J.J. and R.A. Galbraith, Effects of exogenous thyrox-
- ulate the expression of estrogen metabolism enzymes in breast epithelial cells by AhR down-regulation. Mol Cell Biochem, 2017.

- 71. 72. Li, M.Y., et al., Estrogen receptor alpha promotes smoking-carcinogen-induced lung carcinogenesis via cytochrome P450 1B1. *J Mol Med (Berl)*, 2015. **93**(11); p. 1221-33.
- 73. Doostdar, H., M.D. Burke, and R.T. Mayer, Bioflavonoids: selective particle physicochemical properties on toxicological responses of lung cells. J Environ Sci Health A Tox Hazard Subst Environ Eng. 2018 Jaramillo, I.C., et al., Effects of fuel components and combustion
- substrates and inhibitors for cytochrome P450 CYP1A and CYP1B
- Toxicology, 2000. 144(1-3): p. 31-8. Whitten, D.L., et al., The effect of \$1,0hm's wort extracts on CYP3A: a systematic review of prospective clinical trials. Br J Clin Pharmacol. 2006, 62(5): p. 512-26.
- Environ Health Perspect, 1995, 103 Suppl 7: p. 147-50.
 Luckert, C., et al., Polycyclic aromatic hydrocarbons stimulate human CVP3A4 promoter activity via PXR. Toxicol Lett, 2013. 222(2) Bradlow, H.L., et al., Effects of pesticides on the ratio of 16 al-pha/2-hydroxyestrone: a biologic marker of breast cancer risk
- p. 180-8. Wu, W.H., et al., Estrogenic effect of yam ingestion in healthy post menopausal women. *J Am Coll Nutr*, 2005. **24**(4); p. 235-43.
- Dresser, G.K., et al., Evaluation of peppermint oil and ascorbyl palmitate as inhibitors of cytochrome P4503A4 activity in vitro and
- nine antifungal agents and drugs metabolized by human cyto-chromes P450. Curr Drug Metab, 2014. **15**(7): p. 651-79. in vivo. Clin Pharmacol Ther, 2002. **72**(3): p. 247-55. Niwa, T., Y. Imagawa, and H. Yamazaki, Drug interactions between
- tion by estradiol. *Neuropharmacology*, 2003. **45**(7): p. 1011-8. Ho, P.W., et al., Effects of plasticisers and related compounds on Jiang, H., et al., Human catechol-O-methyltransferase down-regula the expression of the soluble form of catechol-O-methyltransferase in MCF-7 cells. *Curr Drug Metab*, 2008. **9**(4): p. 276-9.

<u>%</u>

80. 79. 78. 77. 76. 75. 74.

- 8 % pendent upon gene polymorphisms: a hypothesis. *Med Hypotheses*, 2007, **59**(5); p. 1054-60.

 You'rean, Ma., et al., Phytochemicals inhibit carechol-O-methyltransferase activity in cytosolic fractions from healthy human mammary tissues: implications for catechol estrogen-induced DNA damage. *Transfer 2010* 4 **927** 307 4 **927** 307 4 207 307 6 207 3 Blum, K., et al., Manipulation of catechol-O-methyl-transferase (COMT) activity to influence the attenuation of substance seeking behavior, a subtype of Reward Deficiency Syndrome (RDS), is de-
- damage. Toxicol Sci, 2004. 81(2): p. 316-24.
 Sehiril, A.O., et al., St. John's wort may ameliorate 2,4,6-trinitrobenzenesulfonic add collists of frast through the induction of pregnane.
 X receptors and/or P.glyroproteins. J Physiol Pharmacol, 2015. 66(2):
- 85. Pascussi, J.M., et al., Dexamethasone induces pregnane X recepto and retinoid X receptor-alpha expression in human hepatocytes:
- Ding, X. and J.L. Staudinger, Induction of drug metabolism by for Zhou, H. and P.B. Hylemon, Bile acids are nutrient signaling hormones. Steroids, 2014. **86**: p. 62-8. synergistic increase of CYP3A4 induction by pregnane X receptor activators. *Mol Pharmacol*, 2000. **58**(2): p. 361-72.

skolin: the role of the pregnane X receptor and the protein kinase a signal transduction pathway. *J Pharmacol Exp Ther*, 2005. **312**(2): p. 849-56.

- Mueller, J.W., et al., The Regulation of Steroid Action by Sulfation and Desulfation. *Endocr Rev*, 2015. **36**(5); p. 526-63. Kim, M.S., et al., Suppression of DHEA sulfotransferase (Sult2A1)
- 89. 90. 2004. **287**(4); p. E731-8. Al-Dujaili, E.A., et al., Liquorice and glycyrrhetinic acid increase during the acute-phase response. Am J Physiol Endocrinol Metab
- DHEA and deoxycorticosterone levels in vivo and in vitro by inhibit-ing adrenal SULT2A1 activity. *Mol Cell Endocrinol*, 2011. **336**(1-2): p. 102-9.

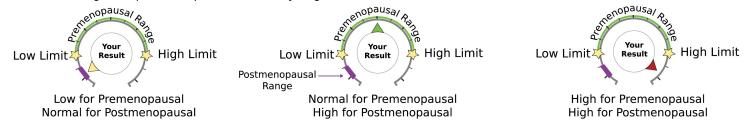
Provider Notes

How to read the DUTCH report

This report is not intended to treat, cure or diagnose any specific diseases. The graphic dutch dials in this report are intended for quick and easy evaluation of which hormones are out of range. Results below the left star are shaded yellow and are below range (left). Results between the stars and shaded green are within the reference range (middle). Results beyond the second star and shaded red are above the reference range (right). Some of these hormones also change with age, and the age-dependent ranges provided should also be considered.



For female reproductive hormones, a purple band is present on the dutch dials. This band represents the expected levels (reference range) for postmenopausal (or non-cycling) women.



In a few places on the graphical pages, you will see fan-style gauges. For sex hormones, you will see one for the balance between 5a/5b metabolism as well as methylation. For adrenal hormones, you will see one to represent the balance between cortisol and cortisone metabolites. These indexes simply look at the ratio of hormones for a preference. An average or "normal" ratio between the two metabolites (or groups of metabolites) will give a result in the middle (as shown here). If the ratio between the metabolites measured is "low" the gauge will lean to the left and similarly to the right if the ratio is higher than normal.

Patient or Sample Comments

Throughout the provider comments you may find some comments specific to your situation or results. These comments will be found in this section or within another section as appropriate. Comments in other sections that are specific to your case will be in **bold**.

The patient reports regular menstrual cycles.

Note: The dates listed on the samples imply that they were older than our allowed 3 weeks when they were received. The instructions ask that patients freeze or refrigerate samples if they are to be held. If that is not the case, the free cortisol and cortisone levels may drop somewhat over time if the samples are too old. Other hormones tested are stable for more than 12 weeks at room temperature. Samples that are refrigerated or frozen are stable for months.

Progesterone Metabolism

The primary role of progesterone is to balance the strong effects of estrogen. Progesterone metabolites are measured and reflect progesterone levels well because very little progesterone is found in urine, so b-Pregnanediol is typically used as a surrogate marker because it is the most abundant metabolite, but we also test the corresponding a-pregnanediol. The average of the two metabolites is reported for progesterone. If levels are in the lower part of the reference range compared to estrogen levels, symptoms of too much estrogen may occur.

When ordering the DUTCH Complete, you will see Progesterone Serum Equivalent on the summary page 1. The urine metabolites of progesterone have been proven to correlate strongly enough to serum progesterone to provide this value. The correlation is the strongest for values within the premenopausal luteal range. Urine metabolites can at times result in somewhat higher serum equivalent results in the postmenopausal range. For this reason the postmenopausal Serum Equivalent range is slightly higher than typical serum ranges. NOTE: If progesterone is taken orally (also with sublingual), these metabolites are elevated from gut metabolism and results do NOT accurately reflect serum levels.

Progesterone levels show she is low for a cycling female in the luteal phase. It is important to check in with her about the timing of her test in relation to her cycle before interpreting this result. If she collected too early or too late, she may have missed her progesterone peak. Ask how she calculated the day to take the samples and when she began her menses after taking the test. If she collected at the right time, this indicates she has not ovulated in the past 5-7 days and may be experiencing anovulatory cycles. She may need support in her HPO axis communication and Chaste Tree berry throughout the month could be considered to help her cycle regularity and ovulation.

Estrogen Metabolism

When evaluating estrogen levels, it is important to assess the following:

• The status (low, normal or high?) of estrogen production:

Levels of the primary ovarian product, estradiol (the strongest estrogen), as well as "total estrogens" may be considered. For women not on HRT, consider the appropriate range (premenopausal or postmenopausal).

• Phase I Metabolism:

Estrogen is metabolized (primarily by the liver) down three phase I pathways. The 2-OH pathway is considered the safest because of the anti-cancer properties of 2-OH metabolites. Conversely, the 4-OH pathway is considered the most genotoxic as its metabolites can create reactive products that damage DNA. The third pathway, 16-OH creates the most estrogenic of the metabolites (although still considerably less estrogenic than estradiol) - 16-OH-E1. If overall estrogen levels are high, production of 16-OH-E1 may exacerbate high estrogen symptoms. Similarly, a woman with very low levels of estrogens, may have less low estrogen symptoms if 16-OH metabolism is preferred. For example Armamento-Villareal showed that a higher 2-OH-E1/16-OH-E1 ratio correlated to bone loss (a low estrogen symptom). Estriol is thought of as a safer (weaker) estrogen metabolite, but it is important to remember that estriol is actually 16-OH-E2, so generally patients that make a lot of the potentially protective/weak estriol may also make a lot of the estrogenic 16-OH-E1.

When evaluating phase I metabolism, it may be important to look at the ratios of the three metabolites to see which pathways are preferred relative to one another. It may also be important to compare these metabolites to the levels of the parent hormones (E1, E2). If the ratios of the three metabolites are favorable but overall levels of metabolites are much lower than E1 and E2, this may imply sluggish phase I clearance of estrogens, which can contribute to high levels of E1 and E2. Similarly, patients with excessive phase I metabolism may have low E1 and E2 levels because of high rates of clearance (as opposed to simply not making a lot of estrogen).

The pie chart will assist you in comparing the three pathway options of phase I metabolism compared to what is "normal." 2-OH metabolism can be increased by using products containing D.I.M. or I-3-C. These compounds are found (or created from) in cruciferous vegetables and are known for promoting this pathway.

Patients typically metabolize a much higher percentage of their estrogens down the more protective 2-OH pathway in phase 1 detoxification. Diindolylmethane (DIM) or Indole-3-Carbinol containing products can help move estrogens more efficiently down this pathway. Be aware that this typically lowers most of the other estrogens, including E1 and E2 as well. If the patients are taking or considering hormone replacement therapy, these products may be considered but a higher dose of estrogen may be needed for the same clinical effect if taken at the same time.

• Methylation (part of phase II metabolism) of estrogens:

After phase I metabolism, both 4-OH and 2-OH (not 16-OH) estrogens can be deactivated and eliminated by methylation. The methylation-activity index shows the patient's ratio of 2-Methoxy-E1 / 2-OH-E1 compared to what is expected. Low methylation can be caused by low levels of nutrients needed for methylation and/or genetic abnormalities (COMT, MTHFR). The COMT enzyme responsible for methylation requires magnesium and methyl donors. Deficiencies in folate or vitamin B6 or B12 can cause low levels of methyl donors. MTHFR genetic defects can make it more difficult for patients to make sufficient methyl donors. Genetic defects in COMT can make methylation poor even in the presence of adequate methyl donors.

Androgen Metabolism

When evaluating androgen levels, it is important to assess the following:

• The status (low, normal or high?) of DHEA:

DHEA and androstenedione are made almost exclusively by the adrenal gland (although a smaller amount is made in the ovaries). These hormones appear in urine as DHEA-S (DHEA-Sulfate), androsterone and etiocholanolone. The best way to assess the total production of DHEA is to add up these three metabolites. This total can be seen on the first page of the DUTCH Complete (and DUTCH Plus). DHEA production decreases quite significantly with age. Age-dependent ranges can be seen on the graphical page of results.

The Total DHEA Production (page 1) was about 2,516ng/mg which is within the overall range but is higher than expected for the patient's age (see the age-dependent ranges). Since these levels are normal for younger individuals, this may not necessarily be a bad thing. High DHEA can cause symptoms of androgen excess including oily skin, acne, sleep problems, headaches and mood disturbances. High levels may be due to supplementation, insulin, stress, elevated prolactin, alcohol and certain medications like ADD meds, Xanax and Wellbutrin. High DHEA can be treated with blood sugar balancing lifestyle, stress reduction and in appropriate cases ashwagandha. In some cases, highly androgenic people may show high levels of both DHEA or testosterone without negative clinical consequence.

Because inflammation blocks DHEA being converted to DHEAS, consider inflammation as a potential part of the overall clinical picture when DHEAS is significantly lower than the downstream metabolites of DHEA (Androsterone, Etiocholanolone) as seen in this case. A sulfur deficiency can also lead to adequate androgens but deficient DHEA-S.

• The status (low, normal or high?) of testosterone:

Females make most of their DHEA in the adrenal gland and a fraction of that DHEA trickles down metabolically to testosterone. For premenopausal women, some testosterone is also made by the ovaries. Levels of testosterone do drop somewhat with age, but not to the degree that DHEA decreases.

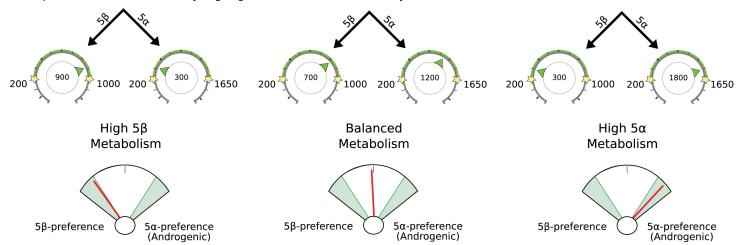
Testosterone levels for this patient were approximately 7.7ng/mg, which is within range. If symptoms

potentially related to high or low testosterone exist, you may also want to carefully evaluate 5a-metabolism (see below). You may also want to evaluate testosterone's downstream metabolites, 5a-androstanediol and 5b-androstanediol. These two metabolites generally parallel testosterone production, although they can also be generated from DHEA without going through testosterone.

• The metabolic preference for the 5a (5-alpha) or 5b (5-beta) pathway:

5a-reductase converts testosterone into 5a-DHT (DHT), which is even more potent (~3x) than testosterone. High levels of DHT can lead to symptoms associated with too much testosterone. Metabolites created down the 5b-pathway are significantly less androgenic than their 5a counterparts. In the examples below, the example on the left shows a patient with 5b-metabolism preference. A patient with a pattern like the example on the right may have high androgen symptoms even though the hormones are in the normal range because of the likely preference for turning a lot of her testosterone into DHT. The fan-style gauge below the hormones shows the 5a or 5b preference based on etiocholanolone (5b) and androsterone (5a) results. Progesterone metabolites are also metabolized by 5a and 5b enzymes and the balance between its two metabolites can be useful to confirm a 5a or 5b preference.

Example of how to read fan-style gauge for 5a-reductase activity:



While testosterone levels are not high, overall DHEA production is on the higher side and androgens are preferring the androgenic 5a pathway. Since the patient did not list significant symptoms of high androgens, these higher levels may be well tolerated by the patient. Since high insulin levels can lead to more DHEA production and 5a-metabolism, it may be worth exploring potential issues with blood sugar and/or insulin.

It is important to consider DHEA and testosterone production, 5a-metabolism patterns as well as the patient symptoms. For example, a woman with higher levels of DHEA and testosterone will often have high androgen symptoms (facial hair, thinning scalp hair, etc.) exacerbated by 5a-metabolism. If, on the other hand, she prefers 5b-metabolism she may not express high androgen symptoms in spite of higher levels of testosterone because 5b is the less androgenic pathway. Testosterone levels may be better understood by also considering its downstream metabolites (5a-androstanediol, 5b-androstanediol). Technically, these metabolites can also be formed from DHEA metabolites without going through the testosterone pathway, but they generally tend to correlate with testosterone production.

You will also see levels of epi-testosterone, which is not androgenic like testosterone. It happens to be produced in about the same concentrations as testosterone (this is an approximate relationship). This can be helpful to assess testosterone therapy and rare cases where testosterone may have other complexities.

DUTCH Adrenal

The HPA-Axis refers to the communication and interaction between the hypothalamus (H) and pituitary (P) in the brain down to the adrenal glands (A) that sit on top of your kidneys. When a physical or psychological stressor occurs, the hypothalamus tells the pituitary to make ACTH, a hormone. ACTH stimulates the adrenal glands to make the stress hormone, cortisol and to a lesser extent DHEA and DHEA-S. Normally, the HPA-axis production follows a daily pattern in which cortisol rises rather rapidly in the first 10-30 minutes after waking (this is the C.A.R.) in order to help with energy, then gradually decreases throughout the day so that it is low at night for sleep. The cycle starts over the next morning. Abnormally high activity occurs in Cushing's Disease where the HPA-axis is hyper-stimulated causing cortisol to be elevated all day. The opposite is known as Addison's Disease, where cortisol is abnormally low because it is not made appropriately in response to ACTH's stimulation. These two conditions are somewhat rare. Examples of more common conditions related to less severely abnormal cortisol levels include fatigue, depression, insomnia, fibromyalgia, anxiety, inflammation and more.

Only a fraction of cortisol is "free" and bioactive. This fraction of cortisol is very important, but levels of metabolized cortisol best represent overall production of cortisol therefore both should be taken into account to correctly assess adrenal function.

When evaluating cortisol levels, it is important to assess the following:

- The overall up-and-down pattern of free cortisol throughout the day, looking for low and high levels: Abnormal results should be considered along with related symptoms.
- The sum of the free cortisol as an expression of the overall tissue cortisol exposure:

This total of five free cortisol measurements is the best way to assess the total of free cortisol throughout the day, but do be aware that it is heavily weighted towards the morning production since three of five measurements are made within the first hour of the day.

• The total level of cortisol metabolites:

We call this calculation "Metabolized Cortisol" which is the sum of a-THF, b-THF and b-THE. While free cortisol is the best assessment for tissue levels of cortisol, it only represents 1-3% of the total produced. The majority of cortisol results in a urine metabolite and the total of these metabolites best represents the total glandular output of cortisol for the day. When overall production is much higher than free cortisol levels, cortisol clearance may be increased (as seen in hyperthyroidism, obesity, etc.) The most common reason for sluggish cortisol clearance (assumed when free cortisol levels are much higher than metabolized cortisol) is low thyroid.

• A potential preference for cortisol or cortisone (the inactive form):

Looking at the comparison between the total for free cortisol and free cortisone is NOT the best indication of a person's preference for cortisol or cortisone. The saliva gland converts cortisol to cortisone in the local tissue. This localized conversion can be seen by comparing cortisol and cortisone levels. To see the patient's preference systemically, it is best to look at which *metabolite* predominates (THF or THE). This preference can be seen in the gauge below metabolized cortisol. This is known as the 11b-HSD index. The enzyme 11b-HSD II converts cortisol to cortisone in the kidneys, saliva gland and colon. 11b-HSD I is more active in the liver, fat cells and the periphery and is responsible for reactivating cortisone to cortisol. Both are then metabolized by 5a-reductase to become tetrahydrocortisol (THF) and tetrahydrocortisone (THE) respectively.

• The Cortisol Awakening Response (CAR):

The unique feature of the DUTCH Plus is the inclusion of the CAR assessment. The response to waking adds one more piece to HPA-axis function. In some cases overall levels of free cortisol may be normal, but the response to stress may be under or overactive. Reasons for a lower CAR might include: an underactive HPA Axis, excessive psychological burnout, seasonal affective disorder (SAD), sleep apnea or poor sleep in general, PTSD, and "chronic fatigue" patients. An elevated CAR can be a result of an over-reactive HPA axis, ongoing job-related stress (anticipatory stress for the day), glycemic dysregulation, pain (ie. waking with painful joints or a migraine), and general depression (not SAD). Scientific literature points to the magnitude of the morning cortisol increase as being connected to HPA-axis health whether the overall production of cortisol is low, normal or high.

- The patient submitted an Insomnia salivary sample. The cortisol result for this sample was 2.10ng/mL. The cortisone result was 10.4 ng/mL. Ranges can be found in the table on the last page.

Nutritional Organic Acids

The following three organic acids are functional markers for vitamin deficiency. These compounds essentially back up in human biochemistry when a key nutrient is missing. These three metabolites have fairly straightforward interpretations. When the markers are elevated, it is likely that the patient's cellular levels of the related nutrient may be insufficient.

Methylmalonate (MMA)

Methylmalonate (also known as methylmalonic acid or MMA) is a functional marker of vitamin B12 (also known as cobalamin) deficiency. When cellular levels of B12 are low either from deficiency or due to a B12 transporter gene mutation, levels of MMA increase. This marker is considered superior to measuring serum B12 levels directly. A 2012 publication by Miller showed that 20% of those tested had a genetic defect in the protein that transports B12 to cells. These patients may have a functional B12 deficiency even if serum levels of B12 are normal.

If levels of MMA are elevated, it may be advisable to increase B12 consumption. Common foods high in B12 include beef liver, sardines, lamb, wild caught salmon, grass-fed beef, nutritional yeast and eggs. Vitamin B12 levels can also be increased through supplementation of B12 (taken as cobalamin, methylcobalamin, hydroxycobalamin, or adenosylcobalamin). Symptoms of a vitamin B12 deficiency include: fatigue, brain fog, memory problems, muscle weakness, unsteady gait, numbness, tingling, depression, migraines/headaches and low blood pressure.

Xanthurenate

Xanthurenate (also known as xanthurenic acid) and Kynurenate (kynurenic acid) are functional markers of vitamin B6 (also known as pyridoxine) deficiency. Vitamin B6 is a critical co-factor to over 100 important reactions that occur in the human body and is stored in the highest concentrations in muscle tissue. Tryptophan is readily converted to NAD by the liver. One of the steps in this pathway requires B6. When there is insufficient B6, xanthurenate is made instead. Kynurenate may also become elevated when patients are B6 deficient because of a different, possibly less B6 dependent pathway. The pathways leading to these biomarkers have other influences, so they will not always agree. When Xanthurenate is elevated, Kynurenate is also elevated about 1/3 of the time. When both are elevated, a B6 deficiency is likely more certain and more severe. Not only is xanthurenate an indicator of a lack of B6, it is also harmful to the human body. It complexes with insulin and decreases insulin sensitivity. In fact, rats fed xanthurenate will actually develop diabetes because of the effects on insulin. If xanthurenate levels are elevated, B6 supplementation may be considered. Food high in B6 include turkey breast, grass-fed beef, pinto beans, avocado, pistachios, chicken, sesame and sunflower seeds.

While there is always some tryptophan going down the kynurenine pathway towards NAD (and possibly xanthurenate), this process is up-regulated by inflammation, estrogen and cortisol. If levels of estrogen or cortisol are high, it may exacerbate xanthurenate elevations and increase the need for B6.

Xanthurenate can also bind to iron and create a complex that increases DNA oxidative damage resulting in higher 8-OHdG levels. If both markers are elevated, there is likely an antioxidant insufficiency.

Xanthurenate and Kynurenate are both elevated in this case, so a vitamin B6 deficiency is likely and may be somewhat significant (since both markers are elevated). It is advisable to consider increasing vitamin B6 intake and to be aware of those things listed above that may induce a vitamin B6 deficiency.

Pyroglutamate

Pyroglutamate (also known as pyroglutamic acid) is a functional marker of glutathione deficiency. Pyroglutamate is a step in the production/recycling of glutathione. If the body cannot convert pyroglutamate forward, it will show up elevated in the urine. High pyroglutamate is an established marker for glutathione deficiency. Pyroglutamate in the urine can also be elevated with cheese consumption.

Glutathione is one of the most potent anti-oxidants in the human body. It is especially important in getting rid of toxins, including the reactive quinone species formed by 4-OH-E1 and 4-OH-E2. This reactive species can damage DNA if not detoxified by either methylation or glutathione.

Some have reported that low pyroglutamate may also be indicative of a need for glutathione; however, this is not established in the scientific literature.

Neurotransmitter Metabolites

The neurotransmitters dopamine, norepinephrine and serotonin are important for human health. Measuring neurotransmitters directly (direct testing of serotonin, for example) is difficult because of their instability and their urinary measurements are controversial with respect to how well they reflect the body's levels of these neuro-hormones. Each of these three neurotransmitters can be assessed indirectly by measuring their urine metabolites. While these metabolites are not a perfect reflection of what's going on in the brain, the scientific literature does affirm their use for a good representation of overall levels of these neurotransmitters.

Homovanillate (HVA)

Homovanillate (also known as HVA) is the primary metabolite of dopamine, a brain and adrenal neurotransmitter that comes from tyrosine (with BH4 and iron as co-factors) and goes on to create norepinephrine (noradrenaline) and epinephrine (adrenaline).

Low levels of HVA can be due to low levels of dopamine or poor conversion of dopamine to HVA. The latter may be due to insufficient levels of SAM, Magnesium, FAD and NAD which are needed to metabolize dopamine. Low circulating dopamine may be due to insufficient BH4, iron or tyrosine. It may also be seen when adrenal function is generally low. Low dopamine levels may be associated with addictions, cravings and pleasure seeking (to boost levels) in addition to sleepiness, impulsivity, tremors, less motivation, fatigue and low mood.

Elevated HVA may be caused by generally increased adrenal hormone output or because of a copper or vitamin C deficiency (which are needed for dopamine conversion to norepinephrine). Elevations may also be caused by a number of medications or supplements including: MAO inhibitors, quercetin, tyrosine, DL-phenylalanine (DLPA), L-dopa, macuna, dopamine medication (Levodopa, Sinemet, Methyldopa), SNRI medication (Wellbutrin), tricyclic antidepressants, amphetamines, appetite suppressants, and caffeine. Bananas also contain dopamine. Elevated dopamine may be associated with loss of memory, insomnia, agitation, hyperactivity, mania, hyper-focus, high stress and anxiety as well as addictions, cravings and pleasure seeking (to maintain high levels).

When HVA is very high, consider if the previously discussed foods, supplements or medications may be the cause. Rarely, tumors associated with increased HVA may be present. In these cases, further testing is necessary for diagnosis. High HVA alone is not diagnostic of a tumor.

Vanilmandelate (VMA)

Vanilmandelate (also known as VMA) is the primary metabolite of norepinephrine and epinephrine (adrenaline). The adrenal gland makes cortisol and DHEA as well as norepinephrine and epinephrine. When adrenal hormone output is generally low, VMA levels may be low. If HVA levels are significantly higher than VMA, there may be a conversion problem from dopamine to norepinephrine. This case can be caused by a copper or vitamin C deficiency. The enzymes COMT (methylation) and MAO are needed to make VMA from norepinephrine. If these enzymes are not working properly, VMA may be low when circulating norepinephrine and/or epinephrine are not low. Low levels of norepinephrine and epinephrine may be associated with addictions, cravings, fatigue, low blood pressure, low muscle tone, intolerance to exercise, depression, loss of alertness. When the body is under physical or psychological stress, VMA levels may increase. Because dopamine gets converted to norepinephrine and ultimately to VMA, the list of medications and supplements that increase HVA may also increase VMA. Elevated levels may be associated with feeling stressed, aggression, violence, impatience, anxiety, panic, worry, insomnia, paranoia, increased tingling/burning, loss of memory, pain sensitivity, high blood pressure and heart palpitations. If VMA and HVA are both extremely high, it may be necessary to rule out a neuroblastic tumor.

Melatonin (measured as 6-OHMS)

Melatonin is not technically an adrenal or sex hormone however it is highly involved in the entire endocrine system. It is made in small amounts in the pineal gland in response to darkness and stimulated by Melanocyte Stimulating Hormone (MSH). A low MSH is associated with insomnia, an increased perception of pain, and mold exposure. Pineal melatonin (melatonin is also made in significant quantities in the gut) is associated with the circadian rhythm of all hormones (including female hormone release). It is also made in small amounts in the bone marrow, lymphocytes, epithelial cells and mast cells. Studies have shown that a urine sample collected upon waking has levels of 6-Hydroxymelatonin-sulfate (6-OHMS) that correlate well to the total levels of melatonin in blood samples taken continuously throughout the night. The DUTCH test uses the waking sample only to test levels of melatonin production.

Low melatonin levels may be associated with insomnia, poor immune response, constipation, weight gain or increased appetite. Elevated melatonin is usually caused by ingestion of melatonin through melatonin supplementation or eating melatonin-containing foods. Elevated melatonin production that is problematic is rare, but levels can be higher in patients with Chronic Fatigue Syndrome and may be phase shifted (peaking later) in some forms of depression.

8-OHdG (8-Hydroxy-2-deoxyguanosine)

8-OHdG (8-hydroxy-2-deoxyguanosine) results can be seen on page 6 of the DUTCH Complete (or DUTCH Plus) report. It is a marker for estimating DNA damage due to oxidative stress (ROS creation). 8-OHdG is considered pro-mutagenic as it is a biomarker for various cancer and degenerative disease initiation and promotion. It can be increased by chronic inflammation, increased cell turnover, chronic stress, hypertension, hyperglycemia/pre-diabetes/diabetes, kidney disease, IBD, chronic skin conditions (psoriasis/eczema), depression, atherosclerosis, chronic liver disease, Parkinson's (increasing levels with worsening stages), Diabetic neuropathy, COPD, bladder cancer, or insomnia. Studies have shown higher levels in patients with breast and prostate cancers. When levels are elevated it may be prudent to eliminate or reduce any causes and increase the consumption of antioxidant containing foods and/or supplements.

The reference range for 8-OHdG is a more aggressive range for Functional Medicine that puts the range limit at the 80th percentile for each gender. A classic range (average plus two standard deviations) would result in a range of 0-6ng/mg for women and 0-10ng/mg for men. Seeking out the cause of oxidative stress may be more crucial if results exceed these limits

Reference Range Determination (last updated 12.20.2018)

We aim to make the reference ranges for our DUTCH tests as clinically appropriate and useful as possible. This includes the testing of thousands of healthy individuals and combing through the data to exclude those that are not considered "healthy" or "normal" with respect to a particular hormone. As an example, we only use a premenopausal woman's data for estrogen range determination if the associated progesterone result is within the luteal range (days 19-21 when progesterone should be at its peak). We exclude women on birth control or with any conditions that may be related to estrogen production. Over time the database of results for reference ranges has grown quite large. This has allowed us to refine some of the ranges to optimize for clinical utility. The manner in which a metabolite's range is determined can be different depending on the nature of the metabolite. For example, it would not make clinical sense to tell a patient they are deficient in the carcinogenic estrogen metabolite, 4-OH-E1 therefore the lower range limit for this metabolite is set to zero for both men and women. Modestly elevated testosterone is associated with unwanted symptoms in women more so than in men, so the high range limit is set at the 80th percentile in women and the 90th percentile for men. Note: the 90th percentile is defined as a result higher than 90% (9 out of 10) of a healthy population.

Classic reference ranges for disease determination are usually calculated by determining the average value and adding and subtracting two standard deviations from the average, which defines 95% of the population as being "normal." When testing cortisol, for example, these types of two standard deviation ranges are effective for determining if a patient might have Addison's (very low cortisol) or Cushing's (very high cortisol) Disease. Our ranges are set more tightly to be optimally used for Functional Medicine practices.

Below you will find a description of the range for each test:

Female Reference Ranges (Updated 06.20.2019)									
	Low%	High%	Low	High		Low%	High%	Low	High
b-Pregnanediol	20%	90%	600	2000	Saliva Cortisol Waking (W)	20%	90%	1.6	4.6
a-Pregnanediol	20%	90%	200	740	Saliva Cortisol (W+30 min.)	20%	90%	3.7	8.2
Estrone (E1)	20%	80%	12	26	Saliva Cortisol (W+60 min.)	20%	90%	2.3	5.3
Estradiol (E2)	20%	80%	1.8	4.5	Saliva Cortisol (Afternoon)	20%	90%	0.4	1.5
Estriol (E3)	20%	80%	5	18	Saliva Cortisol (Night)	0	95%	0	0.9
2-OH-E1	20%	80%	5.1	13.1	Saliva Cortisol (2-3 am)	0	90%	0	0.9
4-OH-E1	0	80%	0	1.8	Saliva Cortisone Waking (W)	20%	90%	6.8	14.5
16-OH-E1	20%	80%	0.7	2.6	Saliva Cortisone (W+30 min.)	20%	90%	12.4	19.4
2-Methoxy-E1	20%	80%	2.5	6.5	Saliva Cortisone (W+60 min.)	20%	90%	9.4	15.3
2-OH-E2	0	80%	0	1.2	Saliva Cortisone Afternoon	20%	90%	2	7.1
4-OH-E2	0	80%	0	0.5	Saliva Cortisone Night	0	95%	0	4.8
2-Methoxy-E2	0	80%	0	0.7	Saliva Cortisone (2-3 am)	0	95%	0	4.8
DHEA-S	20%	90%	20	750	Melatonin (6-OHMS)	20%	90%	10	85
Androsterone	20%	80%	200	1650	8-OHdG	0	90%	0	5.2
Etiocholanolone	20%	80%	200	1000	Methylmalonate	0	90%	0	2.2
Testosterone	20%	80%	2.3	14	Xanthurenate	0	90%	0	1.4
5a-DHT	20%	80%	0	6.6	Kynurenate	0	90%	0	7.3
5a-Androstanediol	20%	80%	12	30	Pyroglutamate	10%	90%	32	60
5b-Androstanediol	20%	80%	20	75	Homovanillate	10%	95%	4	13
Epi-Testosterone	20%	80%	2.3	14	Vanilmandelate	10%	95%	2.4	6.4
a-THF	20%	90%	75	370					
b-THF	20%	90%	1050	2500	Calculated Values				
b-THE	20%	90%	1550	3800	Total DHEA Production	20%	80%	500	3000
% = population perc	antila: Evam	nla - a hiah	limit of 000	% maans	Total Estrogens	20%	80%	35	70
results higher than s			•		Metabolized Cortisol	20%	90%	2750	6500
	will be desid			ejerence	Saliva Cortisol Total	20%	90%	9.6	19.3
range	wiii be uesig	jiiuteu us T	ngn.		Saliva Cortisone Total	20%	90%	36	55

Provider Notes:			



Accession # 00280397 Male Sample Report 123 A Street Sometown, CA 90266



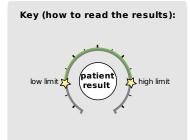
Ordering Physician: Precision Analytical

DOB: 1967-08-09 **Age:** 50

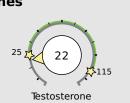
Gender: Male

Collection Times: 2017-08-09 06:01AM (S) 2017-08-09 06:31AM (S) 2017-08-09 07:01AM (S) 2017-08-09 05:01PM (S) 2017-08-09 10:01PM (S) 2017-08-09 06:01AM (U) 2017-08-09 08:01AM (U) 2017-08-09 05:01PM (U) 2017-08-09 05:01PM (U) 2017-08-09 10:01PM (U)

Hormone Testing Summary







 Age
 Range

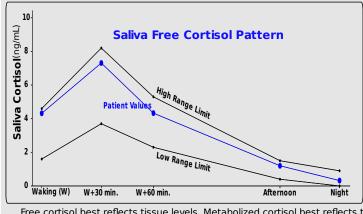
 18-25
 50-115

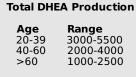
 26-40
 40-95

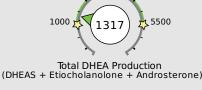
 41-60
 30-80

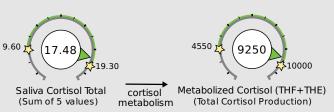
 >60
 25-60

Adrenal Hormones See pages 4 and 5 for a more complete breakdown of adrenal hormones









Free cortisol best reflects tissue levels. Metabolized cortisol best reflects total cortisol production.

The following videos (which can also be found on the website under the listed names along with others) may aid your understanding:

<u>DUTCH Plus Overview</u> (quick overview) <u>Estrogen Tutorial</u> <u>Male Androgen Tutorial</u> <u>Cortisol/CAR Tutorial</u>

PLEASE BE SURE TO READ BELOW FOR ANY SPECIFIC LAB COMMENTS. More detailed comments can be found on page 8.

- The patient collected an "Insomnia" salivary sample in the middle of the night. The cortisol result for this sample was 2.10ng/mL (expected range 0-0.9). Please see page 4 for cortisol and cortisone results for this sample.

The Cortisol Awakening Response (CAR) was 2.99ng/mL (expected range 1.5-4.0) or 69.2% (range 50-160%). See page 5 for more details.



Accession # 00280397 Male Sample Report 123 A Street Sometown, CA 90266



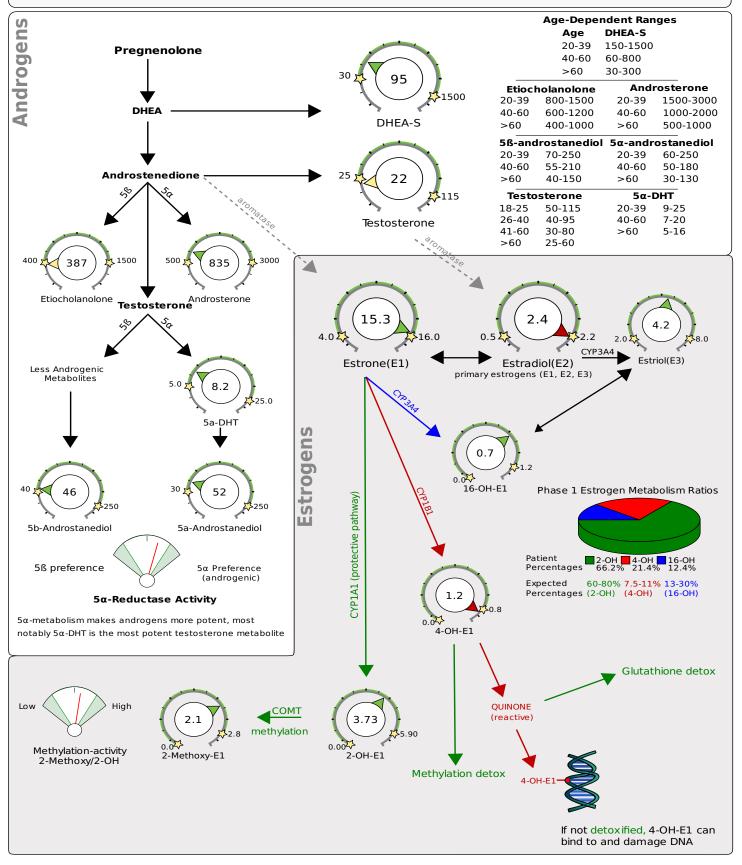
Sex Hormones and Metabolites Ordering Physician:Precision Analytical

DOB: 1967-08-09

Age: 50 Gender: Male Collection Times:
2017-08-09 06:01AM (S)
2017-08-09 06:31AM (S)
2017-08-09 07:01AM (S)
2017-08-09 05:01PM (S)
2017-08-09 10:01PM (S)
2017-08-09 06:01AM (U)
2017-08-09 08:01AM (U)
2017-08-09 05:01PM (U)
2017-08-09 10:01PM (U)

Category	Test		Result	Units	Normal Range
	rone Metabolites (Urine	e)			
	b-Pregnanediol	Low end of range	110.0	ng/mg	75 - 400
	a-Pregnanediol	Low end of range	40.0	ng/mg	20 - 130
Estrogen	s and Metabolites (Urin	e)			
	Estrone(E1)	High end of range	15.3	ng/mg	4 - 16
	Estradiol(E2)	Above range	2.4	ng/mg	0.5 - 2.2
	Estriol(E3)	Within range	4.2	ng/mg	2 - 8
	2-OH-E1	Within range	3.73	ng/mg	0 - 5.9
	4-OH-E1	Above range	1.2	ng/mg	0 - 0.8
	16-OH-E1	Within range	0.7	ng/mg	0 - 1.2
	2-Methoxy-E1	Within range	2.1	ng/mg	0 - 2.8
	2-OH-E2	Above range	0.61	ng/mg	0 - 0.6
	4-OH-E2	Within range	0.1	ng/mg	0 - 0.3
	2-Methoxy-E2	Within range	0.3	ng/mg	0 - 0.8
	Total Estrogen	High end of range	30.34	ng/mg	10 - 34
Androger	ns and Metabolites (Uri	ne)			
	DHEA-S	Low end of range	95.0	ng/mg	30 - 1500
	Androsterone	Low end of range	835.0	ng/mg	500 - 3000
	Etiocholanolone	Below range	387.0	ng/mg	400 - 1500
	Testosterone	Below range	21.6	ng/mg	25 - 115
	5a-DHT	Low end of range	8.2	ng/mg	5 - 25
	5a-Androstanediol	Low end of range	52.0	ng/mg	30 - 250
	5b-Androstanediol	Low end of range	46.0	ng/mg	40 - 250
	Epi-Testosterone	Low end of range	38.1	ng/mg	25 - 115

Hormone metabolite results from the previous page are presented here as they are found in the steroid cascade. See the Provider Comments for more information on how to read the results.





Accession # 00280397 Male Sample Report 123 A Street Sometown, CA 90266

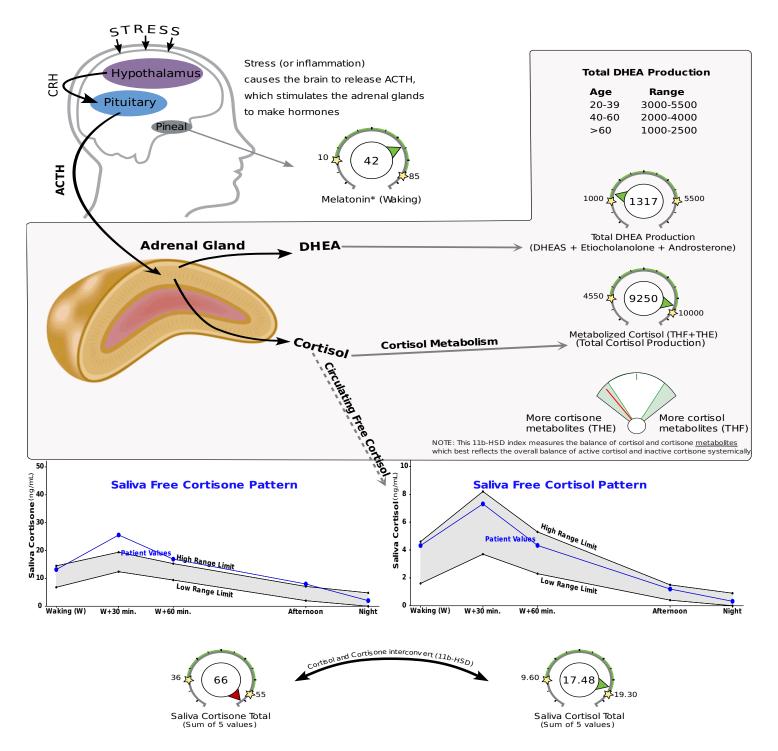


Adrenal Ordering Physician:Precision Analytical

DOB: 1967-08-09

Age: 50 Gender: Male Collection Times:
2017-08-09 06:01AM (S)
2017-08-09 06:31AM (S)
2017-08-09 07:01AM (S)
2017-08-09 05:01PM (S)
2017-08-09 10:01PM (S)
2017-08-09 01:31AM (S*)
2017-08-09 06:01AM (U)
2017-08-09 08:01AM (U)
2017-08-09 05:01PM (U)
2017-08-09 10:01PM (U)

Category	Test		Result	Units	Normal Range
Free Cortis	ol and Cortisone (Saliva)				
	Saliva Cortisol - Waking (W)	High end of range	4.32	ng/mL	1.6 - 4.6
	Saliva Cortisol - W+30 min.	High end of range	7.31	ng/mL	3.7 - 8.2
	Saliva Cortisol - W+60 min.	Within range	4.33	ng/mL	2.3 - 5.3
	Saliva Cortisol - Afternoon	Within range	1.2	ng/mL	0.4 - 1.5
	Saliva Cortisol - Night	Within range	0.32	ng/mL	0 - 0.9
	Saliva Cortisone - Waking (W)	High end of range	13.16	ng/mL	6.8 - 14.5
	Saliva Cortisone - W+30 min.	Above range	25.57	ng/mL	12.4 - 19.4
	Saliva Cortisone - W+60 min.	Above range	16.9	ng/mL	9.4 - 15.3
	Saliva Cortisone - Afternoon	Above range	7.98	ng/mL	2 - 7.1
	Saliva Cortisone - Night	Within range	2.02	ng/mL	0 - 4.8
	Saliva Cortisol Total	High end of range	17.48	ng/mL	9.6 - 19.3
	Saliva Cortisone Total	Above range	65.63	ng/mL	36 - 55
Creatinine	(Urine)				
	Creatinine A (Waking)	Within range	0.45	mg/ml	0.3 - 3
	Creatinine B (Morning)	Within range	0.41	mg/ml	0.3 - 3
	Creatinine C (Afternoon)	Within range	0.9	mg/ml	0.3 - 3
	Creatinine D (Night)	Within range	0.88	mg/ml	0.3 - 3
Cortisol Me	etabolites and DHEA-S (Urine)				
	a-Tetrahydrocortisol (a-THF)	Within range	450.0	ng/mg	175 - 700
	b-Tetrahydrocortisol (b-THF)	Within range	2800.0	ng/mg	1750 - 4000
	b-Tetrahydrocortisone (b-THE)	Above range	6000.0	ng/mg	2350 - 5800
	Metabolized Cortisol (THF+THE)	High end of range	9250.0	ng/mg	4550 - 10000
	DHEA-S	Low end of range	95.0	ng/mg	30 - 1500
Additional	Cortisol and Cortisone (Saliva)				
*	Saliva Cortisol - Insomnia	Above range	2.1	ng/mL	0 - 0.9
*	Saliva Cortisone - Insomnia	Above range	10.4	ng/mL	0 - 4.8



- The patient submitted an Insomnia salivary sample. The cortisol result for this sample was 2.10ng/mL (expected range 0-0.9) The cortisone result for this sample was 10.4 ng/mL (expected range 0-4.8)

The Cortisol Awakening Response (CAR) is the rise in salivary cortisol between the waking sample and the sample collected 30 (as well as 60) minutes later. This "awakening response" is essentially a "mini stress test" and is a useful measurement in addition to the overall up-and-down (diurnal) pattern of free cortisol throughout the day. **This patient shows a waking cortisol of 4.32 and an increase to 7.3 after 30.0 minutes. This is an increase of 2.99ng/mL or 69.2%.** Expected increases differ depending on the methods used. Preliminary research shows that 50-160% or 1.5-4.0ng/mL increases are common with samples collected 30 minutes after waking. These guidelines are considered research only.

This patient shows a salivary cortisol of 4.33 measured 60 minutes after waking. This is an increase of 0.01ng/mL or 0.23% compared to the waking sampe. To date, data suggests that expected results may be 0-70%, and this guideline is considered for research only.



Accession # 00280397
Male Sample Report
123 A Street
Sometown, CA 90266

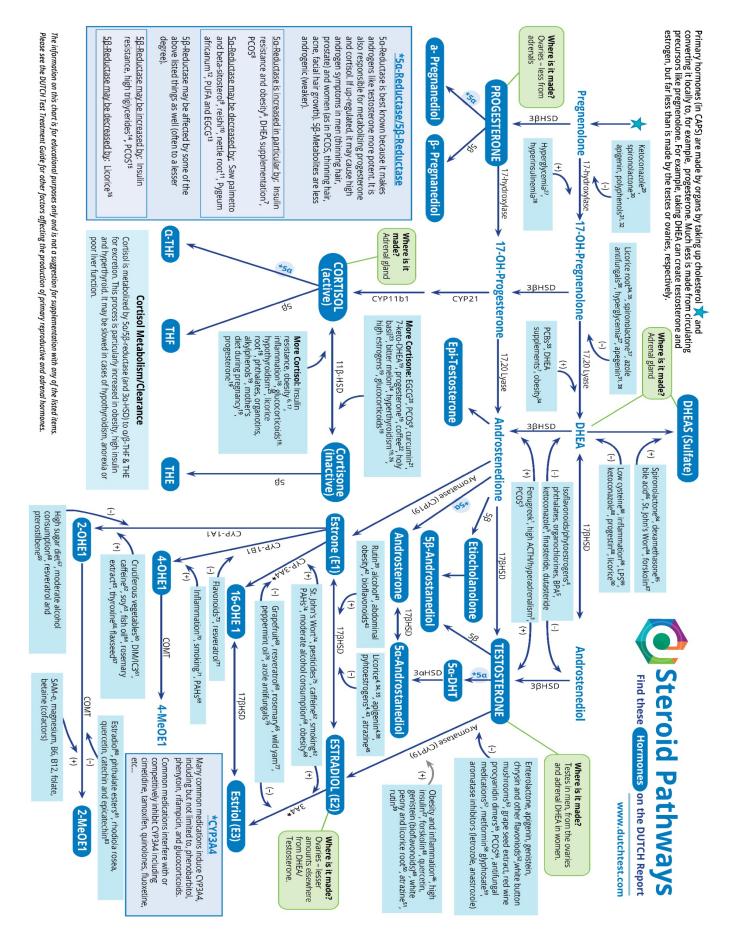


Organic Acid Tests (OATs)
Ordering Physician:
Precision Analytical

DOB: 1967-08-09

Age: 50 Gender: Male Collection Times:
2017-08-09 06:01AM (S)
2017-08-09 06:31AM (S)
2017-08-09 07:01AM (S)
2017-08-09 05:01PM (S)
2017-08-09 10:01PM (S)
2017-08-09 06:01AM (U)
2017-08-09 08:01AM (U)
2017-08-09 05:01PM (U)
2017-08-09 10:01PM (U)

Category	Test		Result	Units	Normal Range				
Nutritional Organic Acids									
Vitamin B12	Marker (may be deficient if high) - (Urine)							
	Methylmalonate (MMA)	Within range	1.2	ug/mg	0 - 3				
Vitamin B6 M	larkers (may be deficient if high) - (Urine)							
	Xanthurenate	Above range	6.8	ug/mg	0 - 2.1				
	Kynurenate	Above range	30.1	ug/mg	0 - 9.3				
Glutathione N	Marker (may be deficient if low o	r high) - (Urine)							
	Pyroglutamate	Below range	23.2	ug/mg	43 - 85				
	Neur	otransmitter Metabo	lites						
Dopamine Mo	etabolite - (Urine)								
	Homovanillate (HVA)	Low end of range	5.6	ug/mg	4.8 - 19				
Norepinephri	ne/Epinephrine Metabolite - (Uri	ine)							
	Vanilmandelate (VMA)	Within range	4.8	ug/mg	2.8 - 8				
Melatonin (*measured as 6-OH-Melatonin-Sulfate) - (Urine)									
	Melatonin* (Waking)	Within range	42.4	ng/mg	10 - 85				
Oxidative Str	Oxidative Stress / DNA Damage, measured as 8-Hydroxy-2-deoxyguanosine (8-OHdG) - (Urine)								
	8-OHdG (Waking)	Within range	4.0	ng/mg	0 - 8.8				



References

Biochem, 2010. **116**(3): p. 146-55. Simonian, M.H., ACTH and thyroid hormone regulation of 3 be-Hamden, K., et al., Potential protective effect on key steroidogen sis and metabolic enzymes and sperm abnormalities by fenugreek steroids in testis and epididymis of surviving diabetic rats. *Arch Physiol*

> 24. 23.

- Kaaijk, E.M., et al., Distribution of steroidogenic enzymes involved cortical cells. J Steroid Biochem, 1986. 25(6): p. 1001-6. ta-hydroxysteroid dehydrogenase activity in human fetal adreno-
- lizing enzymes. J Steroid Biochem Mol Biol, 2005. 93(2-5); p. 285-92. Zhang, S., et al., Endocrine disruptors of inhibiting testicular 38-hydroxysteroid dehydrogenase. Chem Biol Interact, 2019. 303: in androgen synthesis in polycystic ovaries: an immunohistochem cal study. Mol Hum Reprod, 2000. 6(5): p. 443-7. D., et al., Inhibition of 17beta-hydroxysteroid dehydroge-
- dehydrogenase type 1 expression and elevated hepatic 5alpha-re Tomlinson, J.W., et al., Impaired glucose tolerance and insulin resis tance are associated with increased adipose 11beta-hydroxysteroic activity. Diabetes, 2008. 57(10): p. 2652-60.
- Stomati, M., et al., Six-month oral dehydroepiandrosterone supplementation in early and late postmenopause. *Gynecol Endocri* 2000. **14**(5): p. 342-63. ation in early and late postmenopause. Gynecol Endocrino 30. 29.
- rates. Tsilchorozidou, T., J.W. Honour, and G.S. Conway, Altered cortisol metabolism in polycystic ovary syndrome: insulin enhances Salpha-reduction but not the elevated adrenal steroid production J Clin Endocrinol Metab, 2003. **88**(12): p. 5907-13.

<u>ω</u>

- Fujita, R., et al., Anti-androgenic activities of Ganoderma lucidum. of 5-alpha-reductase in the treatment of androgenetic alopecia. J Altern Complement Med, 2002. 8(2): p. 143-52. Prager, $N_{\cdot\cdot}$ et al., A randomized, double-blind, placebo-controlled trial to determine the effectiveness of botanically derived inhibitors 33 32.
- = 10. Moradi, H.R., et al., The histological and histometrical effects of Ethnopharmacol, 2005. 102(1): p. 107-12.
- Urtica dioica extract on rat's prostate hyperplasia. Vet Res Forum, 34
- 2015. 6(1): p. 23-9.
 Will, T., et al., Pygeum africanum for benign prostatic hyperplasia.
 Cochrone Database Syst Rev, 2002(1): p. CD001044.
 Azzouni, F., et al., The S alpha-reductase isozyme family: a review of basic biology and their role in human diseases. Adv Urol, 2012.

36.

35

14. 13 12.

- Metab. 2003. 88(10): p. 4924-31.
 Gambineri, A., et al., Increased clearance of cortisol by 5beta-reduction as in a subgroup of women with adrenal hyperandrogenism in polycystic ovary syndrome. J Endocrinol Invest. 2009. 32(3): p. 210-8.

 Ojima, M., et al., Ilm einhibitory effects of bytycrhizin and bytyr-rheinic acid on the metabolism of cortisol and prednisolone-in 2012: p. 530121.

 Westerbacka, J., et al., Body fat distribution and cortisol metabolism Westerbacka, J., et al., Body fat distribution and cortisol in healthy men: enhanced Speta-reductase and lower cortisol/ in healthy men: enhanced speta-reductase and lower cortisol/ in healthy men: enhanced speta-reductase and lower cortisol/ metabolite ratios in men with fatty liver. J Clin Endocrinol 37. 38.
 - 39
- 40
- 42. Hu, T., et al., Brown adjoose tissue activation by rutin ameliorates polytystic ovary syndrome in rat. J Nutr Biochem, 2017, 47: p. 21-28. Sarkola, T., et al., Actue effect of alcohol on androgens in premenopausal women. Alcohol Alcohol, 2000. 35(1): p. 84-90. Corbould, A.M., et al., The effect of obesity on the ratio of typs 37 (7) beat-hydroxysteroid dehydrogenase mRNA to cytochrome P450 aromatase mRNA in subcutaneous abdominal and nura-abdominal aromatase mRNA in subcutaneous abdominal and nura-abdominal.
- 43. p. 165-75. Krazeisen, A., et al., Human 17beta-hydroxysteroid dehydrogenase adipose tissue of women. Int J Obes Relat Metab Disord, 2002. 26(2)
- type 5 is inhibited by dietary flavonoids. Adv Exp Med Biol, 2002. 505: p. 151-61.
- 4 Le Bail, J.C., et al., Effects of phytoestrogens on aromatase, 3beta breast cancer cells. Life Sci, 2000. 66(14): p. 1281-91. and 17beta-hydroxysteroid dehydrogenase activities and human

ydroxys-

45 Abarikwu, S.O. and E.O. Farombi, Quercetin ameliorates atra-zine-induced changes in the testicular function of rats. *Toxicol* Toxicol Ind

21 22

11β-hydroxysteroid dehydrogenase 1: improving lipid profiles in high-fat-diet-treated rats. PLoS One, 2013. 8(3): p. e49976. Atanasov, A.G., et al., Coffee inhibits the reactivation of glucocorti Hu, G.X., et al., Curcumin as a potent and selective inhibitor of teroid dehydrogenase type 1. PLoS One, 2014. 9(1): p. e84468. catechine-3-gallate, are potent inhibitors of human 11β-h Hintzpeter, J., et al., Green tea and one of its constituents, Epigallo

, et al., Coffee inhibits the reactivation of glucocorti

20. 19.

Chapman, K., M. Holmes, and J. Seckl, 11β-hydroxysteroid dehydro genases: intracellular gate-keepers of tissue glucocorticoid action. *Physiol Rev*, 2013. **93**(3): p. 1139-206.

droxysteroid dehydrogenase type 1 (11β-HSD1) in human adipo-cytes is mediated by MEK, C/EBPβ, and NF-κB/ReIA. J Clin Endocrinol Esteves, C.L., et al., Proinflammatory cytokine induction of 11β-hy activity in subcutaneous adipose tissue in humans: implications in obesity and diabetes. J Clin Endocrinol Metab, 2015. 100(1): p. E70-6 Dube, S., et al., 11β-hydroxysteroid dehydrogenase types 1 and 2

Metab, 2014. 99(1): p. E160-8.

<u>.</u>8 17. 16. 15.

vivo and in vitro studies]. Nihon Naibunpi Gakkai Zasshi, 1990. 66(5)

4.

47. Randolph, J.F., et al., The effect of insulin on aromatase activity in trogens and the molecular underpinnings of aromatase regulation in breast adipose tissue. *Mol Cell Endocrinol*, 2018. **466**: p. 15-30. Health, 2016. **32**(7); p. 1278-85. Gérard, C. and K.A. Brown, Obesity and breast cancer - Role of es-

- Jothle Richard, E., et al., Anti-stress Activity of Ocimum sanctum: Possible Effects on Hypothalamic-Pituitary-Adrenal Axis. *Phytother*
- Res, 2016. 30(5): p. 805-14.

 Blum, A., et al., Momordica chrarantia extract, a herbal remedy for type 2 diabetes, contains a specific 11B-hydroxysteroid dehydrogenase type 1 inhibitor. J Steroid Biochem Mol Biol, 2012. 128(1-2): p.
- tabolites in hyperthyroid and hypothyroid patients. Clin Endocrinol Hoshiro, M., et al., Comprehensive study of urinary cortisol me-
- (0xf), 2006. 64(1); p. 37-45.
 Taniyama, M., K. Honma, and Y. Ban, Urinary cortisol metabolites in the assessment of peripheral thyroid hormone action: application for diagnosis of resistance to thyroid hormone. *Thyroid*, 1993. 3(3):

26. 25.

Eur J Endocrinol, 2002. 146(3): p. 375-80.

Nestler, J.E. and D.J. Jakubowicz, Decreases in ovarian cytochrome and increased 17-hydroxylase activities in type 2 diabetes mellitus Ueshiba, H., et al., Decreased steroidogenic enzyme 17,20-lyase

> 53. 52. 51.

28. 27.

P450c17 alpha activity and serum free testosterone after reduction of insulin secretion in polycystic ovary syndrome. N Engl J Med. 1996. **335**(9): p. 617-23

42

- Endocrinol (Oxf), 1991. 35(2): p. 163-8. Kossor, D.C. and H.D. Colby, Dose-dependent actions of spironolac Engelhardt, D., et al., The influence of ketoconazole on human adrenal steroidogenesis: incubation studies with tissue slices. Clin
- Pharmacology, 1992. **45**(1): p. 27-33.

 Hasegawa, E., et al., Effect of polyphenols on production of steroid tone on the inner and outer zones of the guinea pig adrenal cortex
- Marti, N., et al., Resveratrol inhibits androgen production of human adrenocortical H295R cells by lowering CYP17 and CYP21 expreshormones from human adrenocortical NCI-H295R cells. *Biol Pharm Bull*, 2013. **36**(2): p. 228-37.
- sion and activities. PLoS One. 2017. 12(3): p. e0174224. Andric, S.A., et al., Acute effects of polychlorinated biphenyl-containing and -free transformer fluids on rat testicular steroidogene-
- sis. Environ Health Perspect, 2000. 108(10): p. 955-9.
 Kim, S.H., et al., Body Fat Mass is Associated With Ratio of Steroid Metabolites Reflecting 17.20-Lyase Activity in Perspubertal Girls. J Clin Endocrinol Metab., 2016. 101(12): p. 4653-4660.

60. 59. 58.

- Armanini, D., G. Bonanni, and M. Palermo, Reduction of serum testosterone in men by licorice. N Engl J Med, 1999. 341(15): p. 1158.
 Armanini, D., et al., Licorice reduces serum testosterone in healthy women. Steroids, 2004. **69**(11-12): p. 763-6. Serafini, P. and R.A. Lobo, The effects of spironolactone on adrenal

62. 61.

- steroidogenesis in hirsute women. Fertil Steril. 1985. 44(5): p. 595-9. Ayub. M. and M.J. Levell. Inhibition of human adrenal steroidogenic enzymes in witro by imidazole drugs including ketoconazole. J Steroid Biochem. 1989. 32(4): p. 515-24. Wang. X., et al., Suppression of rat and human androgen biosynthetic enzymes by apligenin: Possible use for the treatment of prostate cancer. Filoteropia, 2016. 111: p. 66-72.
- 63
- 2
- ication enzymes by various extracts of rosemary; description of specific patterns. Food Chem Toxicol, 2001. **39**(9): p. 907-18. Michnovicz, J.J. and R.A. Galbraith, Effects of exogenous thyrox-
- Mahabir, S., et al., Effects of low-to-moderate alcohol supplementa
- ulate the expression of estrogen metabolism enzymes in breast epithelial cells by AhR down-regulation. Mol Cell Biochem, 2017.
- 70. matory cytokines is mediated by the p38 MAP kinase sign duction pathway. Carcinogenesis, 2014. 35(11): p. 2534-43

- 48.
- 49. Watanabe, M. and S. Nakajin, Forskolin up-regulates aromatase (CYP19) activity and gene transcripts in the human adrenocortical carcinoma cell line H295R. J Endocrinol. 2004. 180(1): p. 15-33. Sanderson, J. ..., et al., induction and thibition of aromatase (CYP19) activity by natural and synthetic flavonoid compounds in H295R human adrenocortical carcinoma cells. Toxicol Sci, 2004
- 50. Takeuchi, T., et al., Effect of paeoniflorin, glycyrrhizin and glycyr-rhetic acid on ovarian androgen production. *Am J Chin Med*, 1991
- ity in estrogen sensitive target tissues. J Appl Toxicol, 2008. 28(3): p. 260-70. Holloway, A.C., et al., Atrazine-induced changes in aromatase activ 19(1): p. 73-8.
- Lephart, E.D., Modulation of Aromatase by Phytoestrogens. Enzym Res, 2015. 2015: p. 594656
- Novaes, M.R., et al., The effects of dietary supplementation with Agaricales mushrooms and other medicinal fungi on breast cancer: evidence-based medicine. Clinics (Soo Paulo), 2011. 66(12): p. 2133-
- Satoh, K., et al., Inhibition of aromatase activity by green tea extrac in rats. Food Chem Toxicol, 2002. 40(7): p. 925-33. catechins and their endocrinological effects of oral administration
- p. 8516-22 Eng. E.T., et al., Suppression of estrogen biosynthesis by procyani din dimers in red wine and grape seeds. Cancer Res, 2003. **63**(23)
- Chen, J., et al., The correlation of aromatase activity and obesity in 2015. 8: p. 11. women with or without polycystic ovary syndrome. J Ovarian Res
- Ayub, M. and M.J. Levell, The inhibition of human prostatic aromatase activity by imidazole drugs including ketoconazole and 4-hydroxyandrostenedione. Biochem Pharmacol, 1990. 40(7): p.

57. 56. 55.

- Rice, S., et al., Dual effect of metformin on growth inhibition and oestradiol production in breast cancer cells. *Int J Mol Med*, 2015.
- 35(4): p. 1088-94. Richard. S., et al., Differential effects of glyphosate and roundup on human placental cells and aromatase. *Environ Health Perspect*, 2005. 113(6): p. 716-20.
- Hodges, R.E. and D.M. Minich, Modulation of Metabolic Detoxifi-Scientific Review with Clinical Application. J Nutr Metab, 2015. 2015 cation Pathways Using Foods and Food-Derived Components: A
- Michnovicz, J.J., H. Adlercreutz, and H.L. Bradlow, Changes in levels of urinary estrogen metabolites after oral incide 3-carbinol treat-
- ment in humans. J Natl Concer Inst, 1997. **89**(10): p. 718-23. Sowers, M.R., et al., Selected diet and lifestyle factors are associated.
- women. J Nutr., 2006. 136(6): p. 1588-95.

 Lu, L.J., et al., increased urinary secretion of 2-hydroxyestrone but not 16alpha-hydroxyestrone in premenopausal women during a soya diet containing isoflavones. Cancer Res, 2000. 60(5): p. 1299-305.

87.

- Chen, H.W., et al., The combined effects of garlic oil and fish oil on the hepatic antioxidant and drug-metabolizing enzymes of rats, Br / Nutr, 2003, 89(2); p. 189-200.

 Debersac, P., et al., Induction of cytochrome P450 and/or detox-
- ine on C-2 and C-16 alpha hydroxylations of estradiol in humans Steroids, 1990. 55(1): p. 22-6.
 Peters, L.P., and R.W. Teel. Effect of high sucrose diet on cyto-
- chrome P450 1A and heterocyclic amine mutagenesis. *Anticancer* Res, 2003. **23**(1A): p. 399-403.
- tion on urinary estrogen metabolites in postmenopausal women i a controlled feeding study. Cancer Med, 2017. 6(10): p. 2419-2423. Licznerska, B., et al., Resveratrol and its methoxy derivatives mod-

69. 68 67. 66. 65.

Smerdová, L., et al., Upregulation of CYP1B1 expression by inflam matory cytokines is mediated by the p38 MAP kinase signal trans-

ref 021720

- 71. 72. Li, M.Y., et al., Estrogen receptor alpha promotes smoking-carcinogen-induced lung carcinogenesis via cytochrome P450 1B1. *J Mol Med (Berl)*, 2015. **93**(11); p. 1221-33.
- 73. particle physicochemical properties on toxicological responses of lung cells. J Environ Sci Health A Tox Hazard Subst Environ Eng. 2018 Jaramillo, I.C., et al., Effects of fuel components and combustion
- Doostdar, H., M.D. Burke, and R.T. Mayer, Bioflavonoids: selective substrates and inhibitors for cytochrome P450 CYP1A and CYP1B
- 74. Toxicology, 2000. 144(1-3): p. 31-8. Whitten, D.L., et al., The effect of \$1,0hm's wort extracts on CYP3A: a systematic review of prospective clinical trials. Br J Clin Pharmacol. 2006, 62(5): p. 512-26.
- 76. 75. Environ Health Perspect, 1995, 103 Suppl 7: p. 147-50.
 Luckert, C., et al., Polycyclic aromatic hydrocarbons stimulate human CVP3A4 promoter activity via PXR. Toxicol Lett, 2013. 222(2) Bradlow, H.L., et al., Effects of pesticides on the ratio of 16 al-pha/2-hydroxyestrone: a biologic marker of breast cancer risk
- 78. 77. p. 180-8. Wu, W.H., et al., Estrogenic effect of yam ingestion in healthy post menopausal women. *J Am Coll Nutr*, 2005. **24**(4); p. 235-43.
- Dresser, G.K., et al., Evaluation of peppermint oil and ascorbyl palmitate as inhibitors of cytochrome P4503A4 activity in vitro and in vivo. Clin Pharmacol Ther, 2002. **72**(3): p. 247-55. Niwa, T., Y. Imagawa, and H. Yamazaki, Drug interactions between
- tion by estradiol. *Neuropharmacology*, 2003. **45**(7): p. 1011-8. Ho, P.W., et al., Effects of plasticisers and related compounds on Jiang, H., et al., Human catechol-O-methyltransferase down-regula nine antifungal agents and drugs metabolized by human cyto-chromes P450. Curr Drug Metab, 2014. **15**(7): p. 651-79.

80. 79.

- <u>%</u> % Blum, K., et al., Manipulation of catechol-O-methyl-transferase (COMT) activity to influence the attenuation of substance seeking behavior, a subtype of Reward Deficiency Syndrome (RDS), is dethe expression of the soluble form of catechol-O-methyltransferase in MCF-7 cells. *Curr Drug Metab*, 2008. **9**(4): p. 276-9.
- 8 pendent upon gene polymorphisms: a hypothesis. *Med Hypotheses*, 2007, **59**(5); p. 1054-60.

 You'rean, Ma., et al., Phytochemicals inhibit carechol-O-methyltransferase activity in cytosolic fractions from healthy human mammary tissues: implications for catechol estrogen-induced DNA damage. *Transfer 2010* 4 **927** 307 4 **927** 307 4 207 307 6 207 3
- 84 damage. Toxicol Sci, 2004. 81(2): p. 316-24.
 Sehiril, A.O., et al., St. John's wort may ameliorate 2,4,6-trinitrobenzenesulfonic add collists of frast through the induction of pregnane.
 X receptors and/or P.glyroproteins. J Physiol Pharmacol, 2015. 66(2):
- 86. 85. Pascussi, J.M., et al., Dexamethasone induces pregnane X recepto and retinoid X receptor-alpha expression in human hepatocytes: synergistic increase of CYP3A4 induction by pregnane X receptor activators. *Mol Pharmacol*, 2000. **58**(2): p. 361-72.
- Ding, X. and J.L. Staudinger, Induction of drug metabolism by for Zhou, H. and P.B. Hylemon, Bile acids are nutrient signaling hormones. Steroids, 2014. **86**: p. 62-8. skolin: the role of the pregnane X receptor and the protein kinase a signal transduction pathway. *J Pharmacol Exp Ther*, 2005. **312**(2): p. 849-56.
- Mueller, J.W., et al., The Regulation of Steroid Action by Sulfation and Desulfation. *Endocr Rev*, 2015. **36**(5); p. 526-63. Kim, M.S., et al., Suppression of DHEA sulfotransferase (Sult2A1) during the acute-phase response. Am J Physiol Endocrinol Metab

89. 88

90. 2004. **287**(4); p. E731-8. Al-Dujaili, E.A., et al., Liquorice and glycyrrhetinic acid increase DHEA and deoxycorticosterone levels in vivo and in vitro by inhibit-ing adrenal SULT2A1 activity. *Mol Cell Endocrinol*, 2011. **336**(1-2): p. 102-9.

DutchTest.com

Provider Notes

How to read the DUTCH report

This report is not intended to treat, cure or diagnose any specific diseases. The graphic dutch dials in this report are intended for quick and easy evaluation of which hormones are out of range. Results below the left star are shaded yellow and are below range (left). Results between the stars and shaded green are within the reference range (middle). Results beyond the second star and shaded red are above the reference range (right). Some of these hormones also change with age, and the age-dependent ranges provided should also be considered.



In a few places on the graphical pages, you will see fan-style gauges. For sex hormones, you will see one for the balance between 5a/5b metabolism as well as methylation. For adrenal hormones, you will see one to represent the balance between cortisol and cortisone metabolites. These indexes simply look at the ratio of hormones for a preference. An average or "normal" ratio between the two metabolites (or groups of metabolites) will give a result in the middle (as shown here). If the ratio between the metabolites measured is "low" the gauge will lean to the left and similarly to the right if the ratio is higher than normal.

Patient or Sample Comments

Throughout the provider comments you may find some comments specific to your situation or results. These comments will be found in this section or within another section as appropriate. Comments in other sections that are specific to your case will be in **bold**.

Note: The dates listed on the samples imply that they were older than our allowed 3 weeks when they were received. The instructions ask that patients freeze or refrigerate samples if they are to be held. If that is not the case, the free cortisol and cortisone levels may drop somewhat over time if the samples are too old. Other hormones tested are stable for more than 12 weeks at room temperature. Samples that are refrigerated or frozen are stable for months.

Androgen Metabolism

When evaluating androgen levels, it is important to assess the following:

• The status (low, normal or high?) of DHEA:

DHEA and androstenedione are made almost exclusively by the adrenal gland (although a smaller amount is made in the ovaries). These hormones appear in urine as DHEA-S (DHEA-Sulfate), androsterone and etiocholanolone. The best way to assess the total production of DHEA is to add up these three metabolites. This total can be seen on the first page of the DUTCH Complete (and DUTCH Plus). DHEA production decreases quite significantly with age. Age-dependent ranges can be seen on the graphical page of results.

The Total DHEA Production (page 1) was about 1,317ng/mg which is within the overall range but is below the range for the patient's age-dependent range. This implies that the adrenal glands are not producing appropriate DHEA levels for the patient's age. Low DHEA is associated with depression, diabetes, heart disease, inflammation and immune disorders. It can be caused by hypothyroidism. It can cause fatigue, low mood and low libido. Supplementing DHEA in women often raises both testosterone and estrogen, which may or may not be desirable here. DHEA may increase with adaptogens such as maca and rhodiola, which improve overall adrenal output.

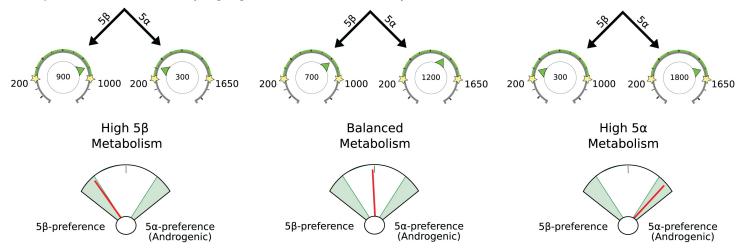
• The status (low, normal or high?) of testosterone:

The testes make most of the male's testosterone. Levels tend to be their highest at around 20 years of age and start to decline when men get into their 30's. Levels continue to drop as men age. Consider the appropriate age-dependent range for your patient. In older men, you can also consider the 18-25 year-old group to approximate what levels may have been when the patient was young and relatively healthy.

• The metabolic preference for the 5a (5-alpha) or 5b (5-beta) pathway:

5a-reductase converts testosterone into 5a-DHT (DHT), which is even more potent (~3x) than testosterone. High levels of DHT can lead to symptoms associated with too much testosterone (thinning scalp hair, acne, etc.) and may also be associated with prostate issues in older men. Metabolites created down the 5b-pathway are significantly less androgenic than their 5a counterparts. In the examples below, the example on the left shows a patient with 5b-metabolism preference. A patient with a pattern like the example on the right may have high androgen symptoms even though testosterone is in the normal range because of the likely preference for turning a lot of his testosterone into DHT. The fan-style gauge below the hormones shows the 5a or 5b preference based on the balance between etiocholanolone (5b) and androsterone (5a) as well as 5a-androstanediol and 5b-androstanediol.

Example of how to read fan-style gauge for 5a-reductase activity:



You will also see levels of epi-testosterone, which is not androgenic like testosterone. It happens to be produced in about the same concentrations as testosterone (this is an approximate relationship). This can be helpful to assess testosterone therapy and rare cases where testosterone may have other complexities.

Estrogen Metabolism

When evaluating estrogen levels, it is important to assess the following:

- The status (low, normal or high?) of estrogen production:
- Levels of the primary estrogen, estradiol (the strongest estrogen), as well as "total estrogens" may be considered.
- Phase I Metabolism:

Estrogen is metabolized (primarily by the liver) down three phase I pathways. The 2-OH pathway is considered the safest because of the anti-cancer properties of 2-OH metabolites. Conversely, the 4-OH pathway is considered the most genotoxic as its metabolites can create reactive products that damage DNA. The third pathway, 16-OH creates the most estrogenic of the metabolites (although still considerably less estrogenic than estradiol) - 16-OH-E1.

When evaluating phase I metabolism, it may be important to look at the ratios of the three metabolites to see which pathways are preferred relative to one another. It may also be important to compare these metabolites to the levels of the parent hormones (E1, E2). If the ratios of the three metabolites are favorable but overall levels of metabolites are much lower than E1 and E2, this may imply sluggish phase I clearance of estrogens, which can contribute to high levels of E1 and E2.

The pie chart will assist you in comparing the three pathway options of phase I metabolism compared to what is "normal." 2-OH metabolism can be increased by using products containing D.I.M. or I-3-C. These compounds are found (or created from) in cruciferous vegetables and are known for promoting this pathway.

• Methylation (part of Phase II Metabolism) of estrogens:

After phase I metabolism, both 4-OH and 2-OH (not 16-OH) estrogens can be deactivated and eliminated by methylation. The methylation-activity index shows the patient's ratio of 2-Methoxy-E1 / 2-OH-E1 compared to what is expected. Low methylation can be caused by low levels of nutrients needed for methylation and/or genetic abnormalities (COMT, MTHFR). The COMT enzyme responsible for methylation requires magnesium and methyl donors. Deficiencies in folate or vitamin B6 or B12 can cause low levels of methyl donors. MTHFR genetic defects can make it more difficult for patients to make sufficient methyl donors. Genetic defects in COMT can make methylation poor even in the presence of adequate methyl donors.

Progesterone levels are of marginal value in men, although deficiency can be associated with some clinical conditions such as depression, fatigue, and low libido.

Progesterone metabolites have limited relevance in male patients, but may be worth considering in some patients with abnormal results. In this case, both progesterone metabolites are within the normal range.

DUTCH Adrenal

The HPA-Axis refers to the communication and interaction between the hypothalamus (H) and pituitary (P) in the brain down to the adrenal glands (A) that sit on top of your kidneys. When a physical or psychological stressor occurs, the hypothalamus tells the pituitary to make ACTH, a hormone. ACTH stimulates the adrenal glands to make the stress hormone, cortisol and to a lesser extent DHEA and DHEA-S. Normally, the HPA-axis production follows a daily pattern in which cortisol rises rather rapidly in the first 10-30 minutes after waking (this is the C.A.R.) in order to help with energy, then gradually decreases throughout the day so that it is low at night for sleep. The cycle starts over the next morning. Abnormally high activity occurs in Cushing's Disease where the HPA-axis is hyper-stimulated causing cortisol to be elevated all day. The opposite is known as Addison's Disease, where cortisol is abnormally low because it is not made appropriately in response to ACTH's stimulation. These two conditions are somewhat rare. Examples of more common conditions related to less severely abnormal cortisol levels include fatigue, depression, insomnia, fibromyalgia, anxiety, inflammation and more.

Only a fraction of cortisol is "free" and bioactive. This fraction of cortisol is very important, but levels of metabolized cortisol

best represent overall production of cortisol therefore both should be taken into account to correctly assess adrenal function.

When evaluating cortisol levels, it is important to assess the following:

- The overall up-and-down pattern of free cortisol throughout the day, looking for low and high levels: Abnormal results should be considered along with related symptoms.
- The sum of the free cortisol as an expression of the overall tissue cortisol exposure:

This total of five free cortisol measurements is the best way to assess the total of free cortisol throughout the day, but do be aware that it is heavily weighted towards the morning production since three of five measurements are made within the first hour of the day.

• The total level of cortisol metabolites:

We call this calculation "Metabolized Cortisol" which is the sum of a-THF, b-THF and b-THE. While free cortisol is the best assessment for tissue levels of cortisol, it only represents 1-3% of the total produced. The majority of cortisol results in a urine metabolite and the total of these metabolites best represents the total glandular output of cortisol for the day. When overall production is much higher than free cortisol levels, cortisol clearance may be increased (as seen in hyperthyroidism, obesity, etc.) The most common reason for sluggish cortisol clearance (assumed when free cortisol levels are much higher than metabolized cortisol) is low thyroid.

• A potential preference for cortisol or cortisone (the inactive form):

Looking at the comparison between the total for free cortisol and free cortisone is NOT the best indication of a person's preference for cortisol or cortisone. The saliva gland converts cortisol to cortisone in the local tissue. This localized conversion can be seen by comparing cortisol and cortisone levels. To see the patient's preference systemically, it is best to look at which *metabolite* predominates (THF or THE). This preference can be seen in the gauge below metabolized cortisol. This is known as the 11b-HSD index. The enzyme 11b-HSD II converts cortisol to cortisone in the kidneys, saliva gland and colon. 11b-HSD I is more active in the liver, fat cells and the periphery and is responsible for reactivating cortisone to cortisol. Both are then metabolized by 5a-reductase to become tetrahydrocortisol (THF) and tetrahydrocortisone (THE) respectively.

• The Cortisol Awakening Response (CAR):

The unique feature of the DUTCH Plus is the inclusion of the CAR assessment. The response to waking adds one more piece to HPA-axis function. In some cases overall levels of free cortisol may be normal, but the response to stress may be under or overactive. Reasons for a lower CAR might include: an underactive HPA Axis, excessive psychological burnout, seasonal affective disorder (SAD), sleep apnea or poor sleep in general, PTSD, and "chronic fatigue" patients. An elevated CAR can be a result of an over-reactive HPA axis, ongoing job-related stress (anticipatory stress for the day), glycemic dysregulation, pain (ie. waking with painful joints or a migraine), and general depression (not SAD). Scientific literature points to the magnitude of the morning cortisol increase as being connected to HPA-axis health whether the overall production of cortisol is low, normal or high.

- The patient submitted an Insomnia salivary sample. The cortisol result for this sample was 2.10ng/mL. The cortisone result was 10.4 ng/mL. Ranges can be found in the table on the last page.

Nutritional Organic Acids

The following three organic acids are functional markers for vitamin deficiency. These compounds essentially back up in human biochemistry when a key nutrient is missing. These three metabolites have fairly straightforward interpretations. When the markers are elevated, it is likely that the patient's cellular levels of the related nutrient may be insufficient.

Methylmalonate (MMA)

Methylmalonate (also known as methylmalonic acid or MMA) is a functional marker of vitamin B12 (also known as cobalamin) deficiency. When cellular levels of B12 are low either from deficiency or due to a B12 transporter gene mutation, levels of MMA increase. This marker is considered superior to measuring serum B12 levels directly. A 2012 publication by Miller showed that 20% of those tested had a genetic defect in the protein that transports B12 to cells. These patients may have a functional B12 deficiency even if serum levels of B12 are normal.

If levels of MMA are elevated, it may be advisable to increase B12 consumption. Common foods high in B12 include beef liver, sardines, lamb, wild caught salmon, grass-fed beef, nutritional yeast and eggs. Vitamin B12 levels can also be increased through supplementation of B12 (taken as cobalamin, methylcobalamin, hydroxycobalamin, or adenosylcobalamin). Symptoms of a vitamin B12 deficiency include: fatigue, brain fog, memory problems, muscle weakness, unsteady gait, numbness, tingling, depression, migraines/headaches and low blood pressure.

Xanthurenate

Xanthurenate (also known as xanthurenic acid) and Kynurenate (kynurenic acid) are functional markers of vitamin B6 (also known as pyridoxine) deficiency. Vitamin B6 is a critical co-factor to over 100 important reactions that occur in the human body and is stored in the highest concentrations in muscle tissue. Tryptophan is readily converted to NAD by the liver. One of the steps in this pathway requires B6. When there is insufficient B6, xanthurenate is made instead. Kynurenate may also become elevated when patients are B6 deficient because of a different, possibly less B6 dependent pathway. The pathways leading to these biomarkers have other influences, so they will not always agree. When Xanthurenate is elevated, Kynurenate is also elevated about 1/3 of the time. When both are elevated, a B6 deficiency is likely more certain and more severe. Not only is xanthurenate an indicator of a lack of B6, it is also harmful to the human body. It complexes with insulin and decreases insulin sensitivity. In fact, rats fed xanthurenate will actually develop diabetes because of the effects on insulin. If xanthurenate levels are elevated, B6 supplementation may be considered. Food high in B6 include turkey breast, grass-fed beef, pinto beans, avocado, pistachios, chicken, sesame and sunflower seeds.

While there is always some tryptophan going down the kynurenine pathway towards NAD (and possibly xanthurenate), this

process is up-regulated by inflammation, estrogen and cortisol. If levels of estrogen or cortisol are high, it may exacerbate xanthurenate elevations and increase the need for B6.

Xanthurenate can also bind to iron and create a complex that increases DNA oxidative damage resulting in higher 8-OHdG levels. If both markers are elevated, there is likely an antioxidant insufficiency.

Xanthurenate and Kynurenate are both elevated in this case, so a vitamin B6 deficiency is likely and may be somewhat significant (since both markers are elevated). It is advisable to consider increasing vitamin B6 intake and to be aware of those things listed above that may induce a vitamin B6 deficiency.

Pyroglutamate

Pyroglutamate (also known as pyroglutamic acid) is a functional marker of glutathione deficiency. Pyroglutamate is a step in the production/recycling of glutathione. If the body cannot convert pyroglutamate forward, it will show up elevated in the urine. High pyroglutamate is an established marker for glutathione deficiency. Pyroglutamate in the urine can also be elevated with cheese consumption.

Glutathione is one of the most potent anti-oxidants in the human body. It is especially important in getting rid of toxins, including the reactive quinone species formed by 4-OH-E1 and 4-OH-E2. This reactive species can damage DNA if not detoxified by either methylation or glutathione.

Some have reported that low pyroglutamate may also be indicative of a need for glutathione; however, this is not established in the scientific literature.

Neurotransmitter Metabolites

The neurotransmitters dopamine, norepinephrine and serotonin are important for human health. Measuring neurotransmitters directly (direct testing of serotonin, for example) is difficult because of their instability and their urinary measurements are controversial with respect to how well they reflect the body's levels of these neuro-hormones. Each of these three neurotransmitters can be assessed indirectly by measuring their urine metabolites. While these metabolites are not a perfect reflection of what's going on in the brain, the scientific literature does affirm their use for a good representation of overall levels of these neurotransmitters.

Homovanillate (HVA)

Homovanillate (also known as HVA) is the primary metabolite of dopamine, a brain and adrenal neurotransmitter that comes from tyrosine (with BH4 and iron as co-factors) and goes on to create norepinephrine (noradrenaline) and epinephrine (adrenaline).

Low levels of HVA can be due to low levels of dopamine or poor conversion of dopamine to HVA. The latter may be due to insufficient levels of SAM, Magnesium, FAD and NAD which are needed to metabolize dopamine. Low circulating dopamine may be due to insufficient BH4, iron or tyrosine. It may also be seen when adrenal function is generally low. Low dopamine levels may be associated with addictions, cravings and pleasure seeking (to boost levels) in addition to sleepiness, impulsivity, tremors, less motivation, fatigue and low mood.

Elevated HVA may be caused by generally increased adrenal hormone output or because of a copper or vitamin C deficiency (which are needed for dopamine conversion to norepinephrine). Elevations may also be caused by a number of medications or supplements including: MAO inhibitors, quercetin, tyrosine, DL-phenylalanine (DLPA), L-dopa, macuna, dopamine medication (Levodopa, Sinemet, Methyldopa), SNRI medication (Wellbutrin), tricyclic antidepressants, amphetamines, appetite suppressants, and caffeine. Bananas also contain dopamine. Elevated dopamine may be associated with loss of memory, insomnia, agitation, hyperactivity, mania, hyper-focus, high stress and anxiety as well as addictions, cravings and pleasure seeking (to maintain high levels).

When HVA is very high, consider if the previously discussed foods, supplements or medications may be the cause. Rarely, tumors associated with increased HVA may be present. In these cases, further testing is necessary for diagnosis. High HVA alone is not diagnostic of a tumor.

Vanilmandelate (VMA)

Vanilmandelate (also known as VMA) is the primary metabolite of norepinephrine and epinephrine (adrenaline). The adrenal gland makes cortisol and DHEA as well as norepinephrine and epinephrine. When adrenal hormone output is generally low, VMA levels may be low. If HVA levels are significantly higher than VMA, there may be a conversion problem from dopamine to norepinephrine. This case can be caused by a copper or vitamin C deficiency. The enzymes COMT (methylation) and MAO are needed to make VMA from norepinephrine. If these enzymes are not working properly, VMA may be low when circulating norepinephrine and/or epinephrine are not low. Low levels of norepinephrine and epinephrine may be associated with addictions, cravings, fatigue, low blood pressure, low muscle tone, intolerance to exercise, depression, loss of alertness. When the body is under physical or psychological stress, VMA levels may increase. Because dopamine gets converted to norepinephrine and ultimately to VMA, the list of medications and supplements that increase HVA may also increase VMA. Elevated levels may be associated with feeling stressed, aggression, violence, impatience, anxiety, panic, worry, insomnia, paranoia, increased tingling/burning, loss of memory, pain sensitivity, high blood pressure and heart palpitations. If VMA and HVA are both extremely high, it may be necessary to rule out a neuroblastic tumor.

Melatonin (measured as 6-OHMS)

Melatonin is not technically an adrenal or sex hormone however it is highly involved in the entire endocrine system. It is made in small amounts in the pineal gland in response to darkness and stimulated by Melanocyte Stimulating Hormone (MSH). A low MSH is associated with insomnia, an increased perception of pain, and mold exposure. Pineal melatonin (melatonin is also made in significant quantities in the gut) is associated with the circadian rhythm of all hormones (including female hormone release). It is also made in small amounts in the bone marrow, lymphocytes, epithelial cells and mast cells. Studies have shown that a urine sample collected upon waking has levels of 6-Hydroxymelatonin-sulfate (6-OHMS) that correlate well to the total levels of melatonin in blood samples taken continuously throughout the night. The DUTCH test uses

the waking sample only to test levels of melatonin production.

Low melatonin levels may be associated with insomnia, poor immune response, constipation, weight gain or increased appetite. Elevated melatonin is usually caused by ingestion of melatonin through melatonin supplementation or eating melatonin-containing foods. Elevated melatonin production that is problematic is rare, but levels can be higher in patients with Chronic Fatigue Syndrome and may be phase shifted (peaking later) in some forms of depression.

8-OHdG (8-Hydroxy-2-deoxyguanosine)

8-OHdG (8-hydroxy-2-deoxyguanosine) results can be seen on page 6 of the DUTCH Complete (or DUTCH Plus) report. It is a marker for estimating DNA damage due to oxidative stress (ROS creation). 8-OHdG is considered pro-mutagenic as it is a biomarker for various cancer and degenerative disease initiation and promotion. It can be increased by chronic inflammation, increased cell turnover, chronic stress, hypertension, hyperglycemia/pre-diabetes/diabetes, kidney disease, IBD, chronic skin conditions (psoriasis/eczema), depression, atherosclerosis, chronic liver disease, Parkinson's (increasing levels with worsening stages), Diabetic neuropathy, COPD, bladder cancer, or insomnia. Studies have shown higher levels in patients with breast and prostate cancers. When levels are elevated it may be prudent to eliminate or reduce any causes and increase the consumption of antioxidant containing foods and/or supplements.

The reference range for 8-OHdG is a more aggressive range for Functional Medicine that puts the range limit at the 80th percentile for each gender. A classic range (average plus two standard deviations) would result in a range of 0-6ng/mg for women and 0-10ng/mg for men. Seeking out the cause of oxidative stress may be more crucial if results exceed these limits.

Reference Range Determination (last updated 12.20.2018)

We aim to make the reference ranges for our DUTCH tests as clinically appropriate and useful as possible. This includes the testing of thousands of healthy individuals and combing through the data to exclude those that are not considered "healthy" or "normal" with respect to a particular hormone. As an example, we only use a premenopausal woman's data for estrogen range determination if the associated progesterone result is within the luteal range (days 19-21 when progesterone should be at its peak). We exclude women on birth control or with any conditions that may be related to estrogen production. Over time the database of results for reference ranges has grown quite large. This has allowed us to refine some of the ranges to optimize for clinical utility. The manner in which a metabolite's range is determined can be different depending on the nature of the metabolite. For example, it would not make clinical sense to tell a patient they are deficient in the carcinogenic estrogen metabolite, 4-OH-E1 therefore the lower range limit for this metabolite is set to zero for both men and women. Modestly elevated testosterone is associated with unwanted symptoms in women more so than in men, so the high range limit is set at the 80th percentile in women and the 90th percentile for men. Note: the 90th percentile is defined as a result higher than 90% (9 out of 10) of a healthy population.

Classic reference ranges for disease determination are usually calculated by determining the average value and adding and subtracting two standard deviations from the average, which defines 95% of the population as being "normal." When testing cortisol, for example, these types of two standard deviation ranges are effective for determining if a patient might have Addison's (very low cortisol) or Cushing's (very high cortisol) Disease. Our ranges are set more tightly to be optimally used for Functional Medicine practices.

Below you will find a description of the range for each test:

	Male Reference Ranges (Updated 12.20.2018)									
	Low%	High%	Low	High		Low%	High%	Low	High	
b-Pregnanediol	10%	90%	75	400	Saliva Cortisol Waking (W)	20%	90%	1.6	4.6	
a-Pregnanediol	10%	90%	20	130	Saliva Cortisol (W+30 min.)	20%	90%	3.7	8.2	
Estrone (E1)	10%	90%	4	16	Saliva Cortisol (W+60 min.)	20%	90%	2.3	5.3	
Estradiol (E2)	10%	90%	0.5	2.2	Saliva Cortisol (Afternoon)	20%	90%	0.4	1.5	
Estriol (E3)	10%	90%	2	8	Saliva Cortisol (Night)	0	95%	0	0.9	
2-OH-E1	0	90%	0	5.9	Saliva Cortisol (2-3 am)	0	90%	0	0.9	
4-OH-E1	0	90%	0	0.8	Saliva Cortisone Waking (W)	20%	90%	6.8	14.5	
16-OH-E1	0	90%	0	1.2	Saliva Cortisone (W+30 min.)	20%	90%	12.4	19.4	
2-Methoxy-E1	0	90%	0	2.8	Saliva Cortisone (W+60 min.)	20%	90%	9.4	15.3	
2-OH-E2	0	90%	0	0.6	Saliva Cortisone Afternoon	20%	90%	2	7.1	
4-OH-E2	0	90%	0	0.3	Saliva Cortisone Night	0	95%	0	4.8	
2-Methoxy-E2	0	90%	0	0.8	Saliva Cortisone (2-3 am)	0	95%	0	4.8	
DHEA-S	20%	90%	30	1500	Melatonin (6-OHMS)	20%	90%	10	85	
Androsterone	20%	80%	500	3000	8-OHdG	0	90%	0	8.8	
Etiocholanolone	20%	80%	400	1500	Methylmalonate	0	90%	0	3	
Testosterone	20%	90%	25	115	Xanthurenate	0	90%	0	2.1	
5a-DHT	20%	90%	5	25	Kynurenate	0	90%	0	9.3	
5a-Androstane diol	20%	90%	30	250	Pyroglutamate	10%	90%	43	85	
5b-Androstanediol	20%	90%	40	250	Homovanillate	10%	95%	4.8	19	
Epi-Testosterone	20%	90%	25	115	Vanilmandelate	10%	95%	2.8	8	
a-THF	20%	90%	175	700						
b-THF	20%	90%	1750	4000	Calculated Values					
b-THE	20%	90%	2350	5800	Total DHEA Production	20%	80%	1000	5500	
% = population perc	ontila, Fyrm	nlo a hi=b	limit of CO	/ magns	Total Estrogens	10%	90%	10	34	
			,		Metabolized Cortisol	20%	90%	4550	10000	
results higher than 909	•	-	,	ence range	Saliva Cortisol Total	20%	90%	9.6	19.3	
W	ll be designa	iea as "nign			Saliva Cortisone Total	20%	90%	36	55	

Provider Notes: